

Effects of Oil Sands Process-Affected Water on Measured Concentrations of Polycyclic Aromatic Hydrocarbons

Hattan A. Alharbi,* Steve B. Wiseman, and John P. Giesy



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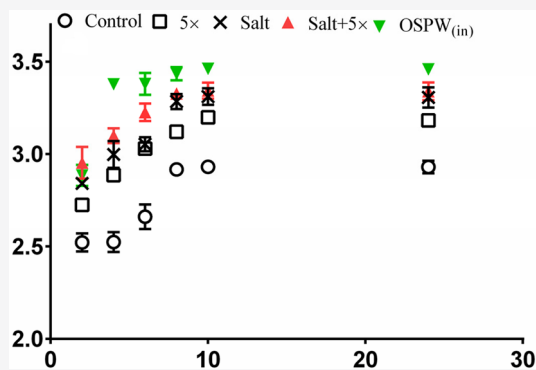
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ABSTRACT: The assessment of the risks and remediation of oil sands process-affected water (OSPW) requires an understanding of possible interactions of naturally occurring surfactants with hydrophobic organic compounds, such as polycyclic aromatic hydrocarbons (PAHs). In this study, the effects of the extractable organic fraction of OSPW (OF-OSPW) on the rate of release and freely dissolved aqueous concentration of several PAHs were investigated using a passive dosing approach, in which a poly-(dimethylsiloxane) (PDMS) film was loaded with the PAHs of interest. The concentrations of PAHs released from PDMS into aqueous solutions were directly proportional to the concentrations of OF-OSPW. PAHs reached equilibrium partitioning 1–4 h faster when the aqueous phase was OF-OSPW than when it was freshwater. Greater concentrations of PAHs were released from PDMS when OF-OSPW was reconstituted in saline water compared to when it was reconstituted in freshwater. The apparent aqueous solubilities of PAHs were greater in intact OSPW than in OF-OSPW. The measured concentrations of PAHs in various aqueous solutions of OSPW and salts correlated with $\log K_{ow}$ of PAHs. This might be because of disruption of PAHs sorbed into PDMS. These results suggest that the bioavailability of PAHs can be greater in the presence of more polar organic compounds in OSPW.

KEYWORDS: OSPW, PAHs, bioavailability, apparent aqueous solubility, PDMS, surfactant, naphthenic acid



1. INTRODUCTION

Extraction of bitumen from surface-mined oil sands deposits in northeastern Alberta, Canada, requires the use of hot, alkaline water to separate bitumen from other unwanted constituents such as clay, salts, and polar organic compounds. This extraction process results in tailing slurries that comprise process-affected water (OSPW), which contains organic and inorganic chemicals, including salts, and solids such as sand and clay that slowly densify to form mature fine tailings (MFTs)¹ that leave behind an aqueous phase of OSPW. One major challenge facing the surface mining oil sands industry is the release of OSPW back into the environment, which has not been done by industry because of a self-imposed policy of zero discharge.² To develop effective strategies for reclamation, remediation, and potential release of OSPW, it is important to understand not only the adverse effects of exposure to OSPW and the chemical composition of OSPW but also the chemodynamics of chemicals in OSPW.^{1,2}

Tailings ponds contain a complex mixture of organic and inorganic chemicals. The aqueous phase of OSPW contains a suite of compounds comprised of not only carbon and hydrogen but also heteroatoms of nitrogen, sulfur, and oxygen (NSO),^{3,4} including naphthenic acids (NAs),¹ as well as salts and metals.⁵ Fine tailings contain nonsoluble organic

contaminants, such as BTX, which includes benzene, toluene, and xylene, humic and fulvic acids, trace metals, asphaltenes, and polycyclic aromatic hydrocarbons (PAHs).⁶ Polycyclic aromatic hydrocarbons make up a group of organic chemicals derived from natural and anthropogenic sources that can be distributed across air, soil, and water bodies.⁷ Polycyclic aromatic hydrocarbons and their metabolites have been detected in sediments of lakes,^{8,9} snowpack,¹⁰ and fish¹¹ near oil sands mining and upgrading operations. Also, oil sands surface mining and processing at bitumen upgrading facilities contribute to atmospheric deposition of PAHs.^{12–17} In oil sands tailing ponds, several PAHs were identified at low concentrations in pore waters.^{18–20} Besides mining and upgrading of bitumen, processes such as a natural erosion of geological formations, forest fires, diesel combustion, residential heating, and airborne dust from roads and mining 60

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61 operations contribute to concentrations of PAHs in the oil
62 sands region.^{15,21}

63 Natural and industrial sources of bitumen can influence
64 chemistries of surface waters and groundwaters in the oil sands
65 region, which might cause mobilization of polar water-soluble
66 bitumen-derived chemicals into the Athabasca River water-
67 shed.^{22–24} The water-soluble fraction (WSF) of bitumen
68 contains NAs, which are surfactant-like, acyclic, monocyclic,
69 and polycyclic acids that are defined by the molecular formula
70 $C_nH_{2n+z}O_2$.¹ Organic acids are the most abundant, water-
71 soluble compounds in bitumen, but their solubilities in water
72 depend on pH.^{25–28} In addition, bitumen and OSPW contain a
73 suite of basic and neutral heterocyclic/aromatic NSO
74 compounds that are soluble at ambient pH.^{4,26,27} OSPW also
75 consists of greater concentrations of inorganic ions such as
76 Na^+ , Cl^- , HCO_3^- , and SO_4^{2-} , which results in the salinity of
77 OSPW being greater than that of natural surface waters of the
78 region.⁵ Because oil sands tailing ponds are a source of PAHs
79 and heteroatomic PAHs, the concentrations of salts and
80 surfactants in tailings ponds might affect the chemodynamics
81 and kinetics of release of PAHs from tailings into the aqueous
82 phase of OSPW.

83 The sequestration of PAHs in soils and sediments is a
84 function of hydrophobicity, and solubility, which is inversely
85 proportional to molecular mass, strong sorption (affinity), and
86 slow release.^{29,30} Lower-mass organic acids, and non-ionic
87 surfactants, can modulate the bioavailability of PAHs by
88 disrupting their sequestration in contaminated soil and
89 sediment and increasing their concentrations in solution.^{31–39}
90 Surfactant-enhanced desorption is one method used to
91 enhance the mobilization, bioavailability, and solubilization of
92 PAHs sorbed to particulates in sediments and soils.⁴⁰ Although
93 enhancement of the bioavailability of PAHs might be
94 important for remediation of PAHs in contaminated sites, it
95 has been argued that it might increase risks to organisms in
96 aquatic environments by increasing the accessible quantity and
97 chemical activity.^{41,42} PAHs and naturally occurring non-ionic
98 and ionic surfactants in bitumen and OSPW might coexist in
99 aquatic environments; however, their interaction is not well
100 understood. Surfactants in OSPW, such as acidic, basic, and
101 neutral, can be released (seepage) into surface waters from
102 deposits of bitumen, and from tailings ponds.^{43,44} PAHs from
103 tailings ponds and bitumen and from the transportation of
104 suspended particles can enter aquatic environments such as
105 surrounding bodies of waters.^{9,10,13}

106 Previously, it has been shown that dissolved organic
107 fractions of OSPW extracted at neutral pH enhance the
108 partitioning of the alkylated PAH, retene, from poly-
109 (dimethylsiloxane) (PDMS) to the aqueous phase, resulting
110 in greater bioavailability to embryos of Japanese medaka
111 (*Oryzias latipes*).⁴⁵ It is therefore possible that aqueous-phase
112 surfactant micelles and salts in OSPW can increase the aqueous
113 concentrations of hydrophobic organic compounds, such as
114 PAHs, by enhancing desorption from suspended fine tailings.
115 The primary aim of this study was to investigate the effects of
116 the extractable organic fraction of OSPW (OF-OSPW), intact
117 OSPW [OSPW_(in)], a commercial mixture of NAs, and salinity
118 on the rates of desorption and dissolved concentrations of
119 PAHs loaded into PDMS. The results of this study could lead
120 to an improved understanding of the fates of PAHs in OSPW
121 and in water containing ionic and non-ionic surfactants from
122 OSPW.

2. MATERIALS AND METHODS

2.1. Chemicals. Acenaphthene (AC), phenanthrene (PH),¹²³
and pyrene (PY) were obtained from Wellington Laboratories¹²⁴
(Guelph, ON). Carbazole (CA), dibenzofuran (DF), dibenzo-¹²⁵
thiophene (DT), fluorene (FL), and a commercial mixture of¹²⁶
NAs were obtained from Sigma-Aldrich (Oakville, ON).¹²⁷
Methanol, ethyl acetate, dichloromethane, and hexane¹²⁸
(HPLC-grade) were purchased from Fisher Scientific¹²⁹
(Edmonton, AB). Acetic acid was purchased from Sigma-¹³⁰
Aldrich. Deionized water was City of Saskatoon municipal¹³¹
water purified using a Milli-Q apparatus. PDMS (aquarium-¹³²
grade sealant) was purchased from Marineland (Blacksburg,¹³³
VA). Reconstituted saltwater (saline solution)⁴⁶ was used to¹³⁴
investigate the influence of salinity on the apparent aqueous¹³⁵
solubility of PAHs. The solution was prepared to mimic the¹³⁶
salinity and major ion concentrations in OSPW. Briefly, 958¹³⁷
mg/L $NaHCO_3$, 663 mg/L $NaCl$, 160 mg/L Na_2SO_4 , 141 mg/¹³⁸
L $CaSO_4 \cdot 2H_2O$, 112 mg/L $MgSO_4 \cdot 7H_2O$, and 16.5 mg/L KCl ¹³⁹
were added to reverse osmosis water.¹⁴⁰

**2.2. Extraction of the Organic Fraction of OSPW (OF-
OSPW).** OSPW was collected from Base Mine Lake in 2015,¹⁴²
which is an end-pit lake constructed from the West-In-Pit¹⁴³
settling basin that received input of tailings from the main¹⁴⁴
extraction facility (Syncrude Canada, Ltd., Fort McMurray,¹⁴⁵
AB). The preparation of OF-OSPW was described pre-¹⁴⁶
viously.²⁸ Briefly, 500 mL of OSPW (pH ~9) was passed¹⁴⁷
through a glass microfiber filter (GF/D 0.47 mm, Whatman)¹⁴⁸
to remove any suspended particulates. Preconcentration of¹⁴⁹
samples was performed in one step using 500 mg of¹⁵⁰
EVOLUTEABN sorbent (Biotage, Charlotte, NC). Before¹⁵¹
the addition of OSPW, the cartridge was conditioned with 6¹⁵²
mL of methanol followed by 6 mL of ultrapure Milli-Q water¹⁵³
(Millipore, Mississauga, ON). Next, 500 mL of OSPW was¹⁵⁴
passed through the cartridge under vacuum at a flow rate of¹⁵⁵
10–15 mL min^{-1} . Subsequently, cartridges were washed with¹⁵⁶
Milli-Q water and allowed to dry under vacuum for 30 min.¹⁵⁷
The total dissolved organic compounds were eluted in one step¹⁵⁸
with 6 mL of methanol, without use of a vacuum. The eluent¹⁵⁹
was evaporated to dryness under a gentle stream of nitrogen¹⁶⁰
and reconstituted in 500 μL of absolute ethanol. Therefore, the¹⁶¹
concentration of dissolved organic compounds in the final¹⁶²
sample was 1000 times greater than in the original sample of¹⁶³
OSPW. The profile of heteroatom classes of this fraction was¹⁶⁴
reported previously.²⁸¹⁶⁵

2.3. Loading of PDMS with PAHs. The aqueous apparent¹⁶⁶
solubility of PAHs in this study was estimated using a¹⁶⁷
partition-controlled delivery system of a PDMS film containing¹⁶⁸
the tested compounds.^{45,47} Briefly, a mixture of PAHs¹⁶⁹
(individual PAHs combined in one mixture) was prepared as¹⁷⁰
a stock solution that was dissolved in a PDMS solution¹⁷¹
prepared in a DCM/hexane solvent (15:85), and the solution¹⁷²
was sonicated until complete dissolution had been achieved.¹⁷³
The final concentration of each PAH in the mixture was 40¹⁷⁴
 $\mu g/mL$. The use of the same procedure yielded a negative¹⁷⁵
control that did not contain PAHs. Next, 3 mL of the mixture¹⁷⁶
was transferred into each 20 mL glass vial, and vials were¹⁷⁷
placed in a fume hood until all solvents evaporated. Vials were¹⁷⁸
wrapped in tin foil, and all procedures were conducted in the¹⁷⁹
dark. Then, 15 mL of ultrapure water (freshwater) was added¹⁸⁰
to each vial. Immediately, vials were spiked with either the¹⁸¹
commercial mixture of NAs at a final concentration of 50 mg/¹⁸²
L, OF-OSPW to give final equivalents of 1X, 2.5X, or 5X OF-¹⁸³

Table 1. Kinetics of the Release into Water of PAHs from PDMS in the Presence of OF-OSPW (1×, 2.5×, and 5×) and a Commercial Mixture of NAs at 50 mg/L^a

	control		1× OF-OSPW		2.5× OF-OSPW		5× OF-OSPW		NAs	
	rate constant (SE)	time for 95% steady state (h)	rate constant (SE)	time for 95% steady state (h)	rate constant (SE)	time for 95% steady state (h)	rate constant (SE)	time for 95% steady state (h)	rate constant (SE)	time for 95% steady state (h)
acenaphthene	0.26 (0.04)	11.35	0.23 (0.04)	12.78	0.30 (0.08)	9.88	0.41 (0.01)	7.20	0.26 (0.04)	11.18
fluorene	0.54 (0.05)	5.48	0.69 (0.05)	4.29	0.58 (0.05)	5.11	0.77 (0.13)	3.87	0.68 (0.06)	4.35
phenanthrene	0.17 (0.03)	17.60	0.17 (0.04)	17.51	0.15 (0.04)	19.12	0.19 (0.02)	15.36	0.17 (0.04)	17.41
pyrene	0.29 (0.04)	10.04	0.28 (0.05)	10.38	0.25 (0.04)	11.80	0.30 (0.02)	9.75	0.28 (0.05)	10.42
carbazole	0.20 (0.02)	14.97	0.20 (0.02)	14.61	0.24 (0.02)	12.08	0.24 (0.03)	12.19	0.24 (0.02)	12.22
dibenzofuran	0.61 (0.05)	4.91	0.59 (0.04)	5.07	0.61 (0.05)	4.84	0.71 (0.10)	4.18	0.58 (0.05)	5.09
dibenzothiophene	0.75 (0.09)	3.97	0.78 (0.09)	3.81	0.64 (0.06)	4.68	0.75 (0.09)	3.98	0.75 (0.06)	3.96

^aValues are the means of three replicates.

184 OSPW, or reconstituted saline water. For OSPW_(in), vials were
 185 filled with filtered OSPW instead of ultrapure water, and the
 186 experiments were conducted in three replicates. One milliliter
 187 of solution was sampled from each vial at 0, 2, 4, 6, 8, 10, and
 188 24 h and stored at −4 °C until analysis.

189 **2.4. Quantification of PAHs in Solution.** Aqueous
 190 concentrations of PAHs were measured in samples taken at
 191 0, 2, 4, 6, 8, 10, and 24 h in the solvent control, OF-OSPW,
 192 OSPW_(in), and solution of salts. Extraction of PAHs was
 193 performed using a liquid–liquid method.⁴⁵ Briefly, 1 mL of the
 194 aqueous solution at each time point was extracted twice with
 195 0.5 mL of a 1:1 hexane/ethyl acetate mixture. Vials were
 196 vortexed and centrifuged at 9000g for 5 min and concentrated
 197 under a gentle stream of nitrogen. Next, samples were
 198 reconstituted in 100 μL of a nonane/hexane mixture (15:85)
 199 and transferred to an autosampler vial fitted with a glass insert.
 200 The concentrations of PAHs were determined using an Agilent
 201 (Santa Clara, CA) model 7890A gas chromatograph system
 202 coupled to an Agilent model 5975C mass spectrometer
 203 operated in electron impact ionization mode (EI) with an
 204 energy of 70 eV. Using a splitless mode, samples were injected
 205 at a volume of 1 μL at an injection port temperature of 280 °C.
 206 Chromatographic separation of PAHs was achieved by use of a
 207 DB-5MS [60 m × 0.25 mm (inside diameter) × 0.25 μm (film
 208 thickness)] fused silica capillary GC column (Agilent). Helium
 209 was used as a carrier gas at a constant flow rate of 1.5 mL
 210 min^{−1}. The following GC oven temperature program was used:
 211 100 °C for 1 min, increase at a rate of 5 °C min^{−1} to 190 °C
 212 for 2 min, increase at a rate of 20 °C min^{−1} to 220 °C for 2
 213 min, and increase at a rate of 40 °C min^{−1} to 300 °C for 4 min.
 214 The GC/MS transfer line was maintained at 28 °C. The
 215 analysis was performed using selected ion monitoring (SIM),
 216 with ions at *m/z* 166 for fluorene, *m/z* 167 for carbazole, *m/z*
 217 168 for dibenzofuran, *m/z* 184 for dibenzothiophene, *m/z* 153
 218 for acenaphthene, *m/z* 178 for phenanthrene, and *m/z* 202 for
 219 pyrene. The limit of detection range was 0.5–2.0 μg/L for
 220 PAHs. The concentrations of targeted PAHs in OF-OSPW
 221 were lower than the limit of detection.

222 **2.5. Data Analysis.** The release of PAHs into different
 223 aqueous phases was analyzed using a one-compartment
 224 model⁴⁸ (eq 1)

$$225 \quad C_{\text{Water}(t)} = C_{\text{Water}(eq)}(1 - \exp^{-k_{\text{Release}}t}) \quad (1)$$

226 where $C_{\text{Water}(t)}$ (milligrams per liter) is the concentration in
 227 water at time t , $C_{\text{Water}(eq)}$ (milligrams per liter) is the
 228 concentration in water at equilibrium, and k_{Release} (inverse

hours) is the release rate constant. Data were fitted using the
 least-squares method using GraphPad Prism 6.07 (GraphPad,
 San Diego, CA), giving estimates of the appropriate rate
 constant and equilibrium concentrations. The time to reach
 95% steady state was used to quantify the time to reach
 equilibrium (eq 2).

$$235 \quad t_{95\% \text{ ss}} = 3/k \quad (2)$$

where 95% ss (hours) is the time to reach 95% of the steady
 state and k is the release rate constant.

The increment or enhancement ratio (r)³⁴ was used to
 describe the effects of treatments on the measured
 concentrations of targeted PAHs in the aqueous phase. Greater
 values of r show a stronger tendency of treatment to enhance
 the release of PAHs from PDMS into the aqueous phase. The
 value of r of each PAH from PDMS was calculated (eq 3).

$$243 \quad r (\%) = (C_b - C_a)/C_a \times 100 \quad (3)$$

where C_a is the desorbed amount of PAH in the control and C_b
 is the desorbed amount of PAH in treatments. The relationship
 of PAH increment ratio to $\log K_{ow}$ was tested by linear
 regression; a p value of <0.05 was considered statistically
 significant.

3. RESULTS AND DISCUSSION

**3.1. Kinetics of the Release of PAHs into a Solution of
 OF-OSPW and Commercial NAs.** Equilibrium concentra-
 tions of each PAH, including acenaphthene, fluorene,
 phenanthrene, pyrene, carbazole, dibenzofuran, and dibenzo-
 thiophene, from PDMS to the freshwater control, OF-OSPW,
 and a commercial mixture of NAs under static conditions were
 reached in 4–18 h (Table 1). In previous studies, the
 concentrations of PAHs released from PDMS into water
 reached equilibrium within 24 h under static conditions.
 The constants for the rate of release and the time to reach a
 steady state for acenaphthene, fluorene, phenanthrene, pyrene,
 carbazole, dibenzofuran, and dibenzothiophene in each of the
 test solutions are summarized in Table 1. Compared to the
 freshwater control, the time for PAHs to reach 95% steady
 state decreased in the presence of OF-OSPW but not NAs at
 50 mg/L. OSPW contains a complex mixture of naturally
 occurring acidic and non-acidic heteroatom-containing organic
 compounds that have surfactant properties.^{3,49} Although
 commercial NAs are not representative of the acid extractable
 fraction of OSPW, it was tested here to determine if complex
 mixtures of carboxylic acids enhance the desorption of PAHs.

271 Compared to OF-OSPW, NAs did not enhance desorption of
 272 PAHs. This is probably because OF-OSPW is a complex
 273 mixture of anionic, cationic, and non-ionic chemicals
 274 representing a mixture of diverse structures, molecular masses,
 275 and physicochemical properties, while commercial NAs
 276 contain only carboxylic acids. At 1× and 2.5× OF-OSPW,
 277 there were no changes in equilibrium concentrations compared
 278 to the control. When the concentration of OF-OSPW was
 279 increased to 5×, the times to reach the 95% steady state of
 280 acenaphthene, fluorene, phenanthrene, pyrene, carbazole, and
 281 dibenzofuran decreased by 4.15, 1.61, 2.24, 0.26, 2.78, and 0.73
 282 h, respectively, compared to the control (Table 1), but the
 283 time for dibenzothiophene to reach equilibrium did not
 284 change.

285 The release of PAHs from PDMS into each aqueous
 286 solution over the course of 24 h is shown (Figures 1–3). The
 287 concentrations of acenaphthene, fluorene, phenanthrene,
 288 pyrene, carbazole, dibenzofuran, and dibenzothiophene in
 289 OF-OSPW increased with time. The greatest calculated r when
 290 the aqueous solution was 1× OF-OSPW was 10% for
 291 acenaphthene and carbazole, while other PAHs had values of
 292 r of <10% (Figure 4A). Values of r , when the aqueous phase
 293 was 2.5× OF-OSPW, were 24%, 20%, 40%, 39%, 20%, 24%,
 294 and 17% for acenaphthene, fluorene, phenanthrene, pyrene,
 295 carbazole, dibenzofuran, and dibenzothiophene, respectively
 296 (Figure 4B). Values of r of acenaphthene, fluorene,
 297 phenanthrene, pyrene, carbazole, dibenzofuran, and dibenzo-
 298 thiophene increased by 51%, 38%, 78%, 86%, 25%, 67%, and
 299 16%, respectively, in 5× OF-OSPW compared to the
 300 freshwater control (Figure 4C). The low solubility and high
 301 affinity of PAHs for soil or sediment decrease their
 302 bioavailability to aquatic organisms. The results of this study
 303 showed that dissolved organic compounds in OSPW can
 304 increase the concentrations and solubilities of PAHs, as shown
 305 previously with retene.⁴⁵ The rates of desorption of
 306 acenaphthene, dibenzofuran, dibenzothiophene, and fluorene
 307 were stable during the sampling period, when the commercial
 308 mixture of NAs was the aqueous phase. However, the
 309 measured concentrations of PAHs did not increase. Values of
 310 r of acenaphthene, fluorene, phenanthrene, pyrene, carbazole,
 311 dibenzofuran, and dibenzothiophene from PDMS incubated
 312 with the commercial mixture of NAs changed by 5%, 2%, 8%,
 313 8%, -8%, 13%, and 3%, respectively (Figure 4D). The effects
 314 of OF-OSPW on the rate of desorption are likely caused by the
 315 addition of non-ionic and ionic surfactants at concentrations
 316 greater than its critical micelle concentration (CMC), which
 317 has been shown to enhance the solubility, rate of release, and
 318 bioavailability of slowly desorbing PAHs in soil water.^{31,32,35,39}

319 Solutions of a mixture of surfactants increase the solubilization
 320 capacity more than solutions of a single surfactant.
 321 Furthermore, the desorption capacity of mixtures of anionic
 322 and non-ionic surfactants is greater than the desorption
 323 capacity of mixtures of anionic and cationic surfactants or
 324 cationic and non-ionic surfactants.⁵⁰

325 The desorption and apparent aqueous solubility of PAHs
 326 increased as the concentration of dissolved organics from
 327 OSPW increased. When the concentrations of surfactants are
 328 lower than the CMC, surfactants occur as individual molecules
 329 or monomers and have minimal effects on the solubilities of
 330 hydrophobic compounds.⁵¹ Thus, the concentrations of
 331 organic compounds in 1× OF-OSPW might not have been
 332 sufficient for self-aggregation or formation of micelles that are
 333 needed to promote partitioning of PAHs from PDMS.

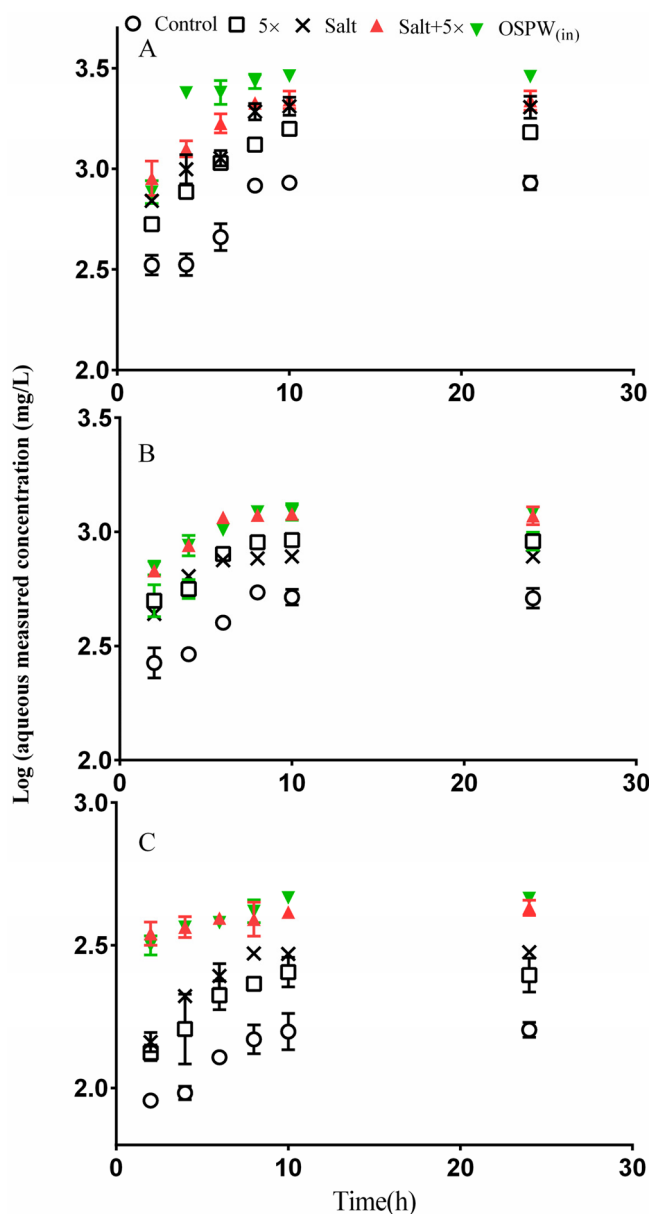


Figure 1. Desorption kinetics of (A) phenanthrene, (B) acenaphthene, and (C) pyrene released from PDMS. PAHs were loaded into PDMS and incubated with 5× OF-OSPW, salts, 5× OF-OSPW+salts, and OSPW_(in). Values are the means of three replicates.

Although heterocyclic PAHs (NSO) are more polar and
 more soluble than nonpolar PAHs (homocyclic) because of
 substitution of one carbon atom with nitrogen, sulfur, or
 oxygen,⁵² the concentrations of carbazole, dibenzofuran, and
 dibenzothiophene in either of the solutions were not greater
 than those of non-NSO PAHs. The concentrations of dissolved
 organic compounds are greater in fresh OSPW than in aged
 OSPW.⁵³ Thus, it is expected that micelle formation is more
 likely to occur in fresh OSPW than in aged OSPW, and that
 the potential for PAHs to partition to the aqueous phase
 decreases as OSPW in tailings ponds is diluted and dissolved
 organics are biodegraded.^{39,53}

The organic compounds extracted from OSPW can vary at
 different pH values,^{25,28} which might influence the desorption
 of PAHs from PDMS to the aqueous phase. It is not known
 whether the observed effect of OF-OSPW on the solubilities of

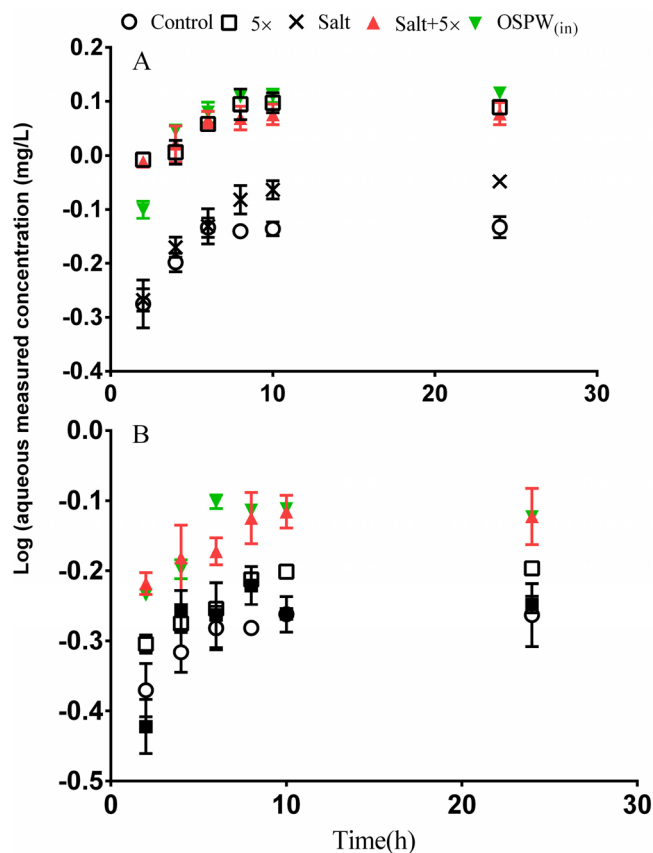


Figure 2. Desorption kinetics of (A) dibenzofuran and (B) dibenzothiophene released from PDMS. PAHs were loaded into PDMS and incubated with 5× OF-OSPW, salts, 5× OF-OSPW+salts, and OSPW_(in). Values are the means of three replicates.

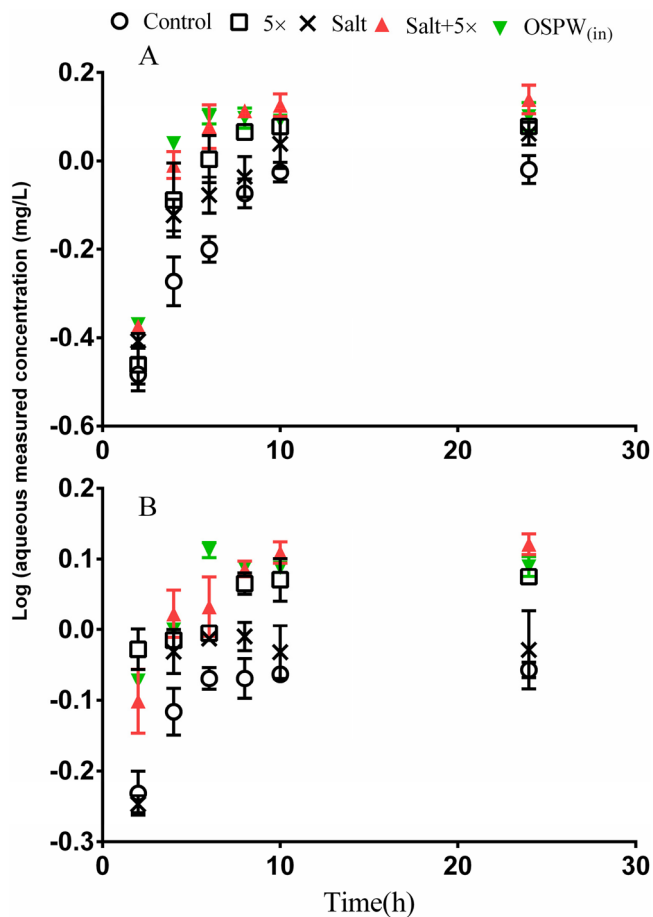


Figure 3. Desorption kinetics of (A) carbazole and (B) fluorene released from PDMS. PAHs were loaded into PDMS and incubated with 5× OF-OSPW, salts, 5× OF-OSPW+salts, and OSPW_(in). Values are the means of three replicates.

350 PAHs was due to concentrations of anionic, cationic, or non-
 351 ionic surfactants. It has been shown that the pH of OSPW is in
 352 the range of 7.7–8.6,⁵⁴ which is close to the pH of the total
 353 organic extract used in this study. Although in OSPW the
 354 concentrations of acid compounds are greater than the
 355 concentrations of basic and neutral compounds, it is expected
 356 that the extraction method used in this study resulted in
 357 concentrations of basic and neutral compounds being greater
 358 than the concentration of organic acids in OF-OSPW.^{25,28}
 359 Therefore, the enhancement of the solubilities of PAHs might
 360 be caused by the mixture of dissolved organic compounds (i.e.,
 361 acidic, basic, and neutral) in OSPW. It has been shown that
 362 excess concentrations of organic acids in soil water might
 363 decrease the efficiency to dissociate hydrophobic compounds
 364 from soil.³⁶ Given the differences in the chemical structures
 365 and properties of organic compounds in OSPW, the extraction
 366 of OSPW at pH values similar to those of tailing ponds will
 367 provide an environmentally relevant representation of the total
 368 organic fraction. The desorption efficiency of PAHs is strongly
 369 related to the dissociation forms of organic acids (more ions)
 370 in solution, which can be achieved at higher pH values.³⁶
 371 **3.2. Effects of OSPW_(in) and Salts on the Kinetics of**
 372 **Release of PAHs.** The kinetics of desorption of PAHs from
 373 PDMS were influenced by salts, which are present at relatively
 374 high concentrations within OSPW.⁵ The desorption of PAHs
 375 from PDMS differed depending on whether the aqueous phase
 376 consisted of salts, salts mixed with 5× OF-OSPW, or OSPW_(in)
 377 (Figures 1–3). When the aqueous phase consisted of salts

dissolved in freshwater, the measured concentrations of PAHs
 378 were slightly increased. The times for PAHs to reach a steady
 379 state (equilibrium) when the aqueous phase consisted of salts,
 380 salts with 5× OF-OSPW, and OSPW_(in) are listed in Table 2.
 381 Equilibria of freely dissolved concentrations of acenaphthene,
 382 fluorene, phenanthrene, pyrene, carbazole, dibenzofuran, and
 383 dibenzothiophene were reached either faster or slower than for
 384 the control. For example, pyrene reached 95% of steady state
 385 faster by ~7 h compared to the control in salts with 5× OF-
 386 OSPW, while dibenzofuran was slower by approximately 2 h,
 387 reaching a steady state in salts compared to the freshwater
 388 control (Table 2).
 389

Values of *r* when the aqueous phase consisted of salts in
 390 freshwater were 77%, 15%, 138%, 56%, 20%, 21%, and 3% for
 391 acenaphthene, fluorene, phenanthrene, pyrene, carbazole,
 392 dibenzofuran, and dibenzothiophene, respectively (Figure 5).
 393 The rate of release of PAHs from PDMS into the solution of
 394 salts with 5× OF-OSPW was greater than the rate of release
 395 into salts or 5× OSPW alone. Values of *r* when the aqueous
 396 solution consisted of salts with 5× OSPW were 129%, 50%,
 397 157%, 167%, 44%, 62%, and 38% for acenaphthene, fluorene,
 398 phenanthrene, pyrene, carbazole, dibenzofuran, and dibenzo-
 399 thiophene, respectively (Figure 5). The desorption efficiency
 400 of PAHs was enhanced in the presence of OSPW_(in), and
 401 values of *r* for acenaphthene, fluorene, phenanthrene, pyrene,
 402 carbazole, dibenzofuran, and dibenzothiophene were 136%, 403

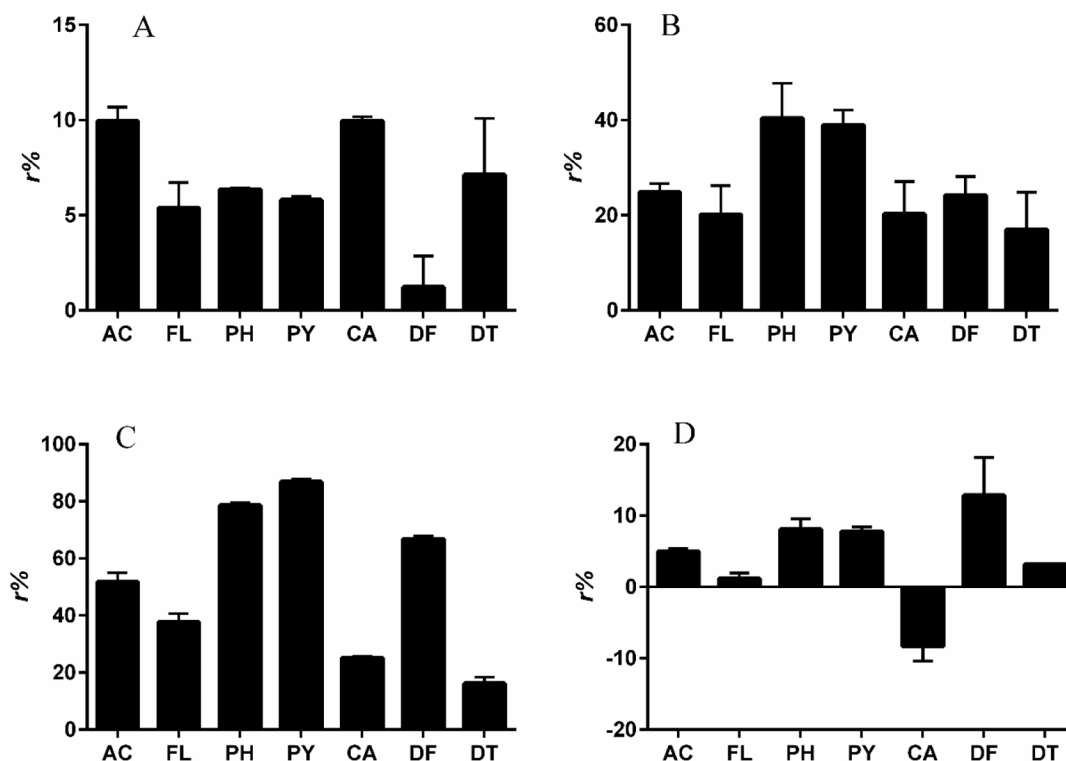


Figure 4. Effect of OF-OSPW on the increment ratio (r) of PAHs. The effects of (A) 1× OF-OSPW, (B) 2.5× OF-OSPW, (C) 5× OF-OSPW, and (D) NAs on the release of PAHs from PDMS to a water solution measured once concentrations reached a steady state (see the calculation of r in Data Analysis).

Table 2. Kinetics of Release into Water of PAHs from PDMS in the Presence of Salts, Salts with 5× OF-OSPW and OSPW_(in)^a

	control		salts		salts with 5× OF-OSPW		OSPW _(in)	
	rate constant (SE)	time for 95% steady state (h)	rate constant (SE)	time for 95% steady state (h)	rate constant (SE)	time for 95% steady state (h)	rate constant (SE)	time for 95% steady state (h)
acenaphthene	0.26 (0.04)	11.35	0.31 (0.06)	9.51	0.39 (0.05)	7.50	0.42 (0.06)	7.05
fluorene	0.54 (0.05)	5.48	0.53 (0.09)	5.60	0.43 (0.05)	6.83	0.53 (0.05)	5.63
phenanthrene	0.17 (0.03)	17.60	0.17 (0.03)	16.72	0.23 (0.03)	12.64	0.27 (0.04)	11.00
pyrene	0.29 (0.04)	10.04	0.30 (0.05)	9.68	0.92 (0.17)	3.25	0.53 (0.07)	5.58
carbazole	0.20 (0.02)	14.97	0.21 (0.02)	13.86	0.25 (0.03)	11.54	0.33 (0.54)	9.08
dibenzofuran	0.61 (0.05)	4.91	0.41 (0.04)	7.16	0.81 (0.09)	3.66	0.47 (0.01)	6.38
dibenzothiophene	0.75 (0.09)	3.97	0.59 (0.07)	5.07	0.78 (0.12)	3.79	0.64 (0.06)	4.67

^aValues are the means of three replicates.

404 40%, 237%, 188%, 32%, 77%, and 37%, respectively (Figure 5).
 405 The solubilization of PAHs by surfactants can be enhanced in
 406 saline solutions. It has been shown that mixed micelles of
 407 surfactants were more stable in the presence of salts, and the
 408 solubility of pyrene and desorption from soil in mixtures of
 409 saline anionic and non-ionic surfactants were greater than in a
 410 nonsaline solution.³⁵ Although salts in OSPW have been
 411 shown to be toxic to aquatic organisms,⁴⁶ the effects of salts in
 412 OSPW on solubility enhancement of PAHs have never been
 413 investigated. The concentrations of Na⁺, Cl⁻, and HCO₃³⁻ in
 414 OSPW from Base Mine Lake are greater than in freshwater but
 415 lower than in pore water of fluid fine tailings (FFT).⁵⁵ The
 416 concentrations of PAHs were greater when the aqueous phase
 417 was a mixture of salts and a 5× OF-OSPW solution compared
 418 to concentrations in either OF-OSPW or the saline solution,
 419 alone. Additionally, the increment ratio of desorption of PAHs
 420 was almost identical when the aqueous phase consisted of
 421 OSPW_(in) or salts with 5× OF-OSPW. Because the organic
 422 compounds and inorganic ions coexist in OSPW, these

observations suggest that the salinity of OSPW is a critical
 component affecting desorption of PAHs from tailings by
 surfactants.

3.3. Correlations between the Enhancement Ratio and K_{ow} of Targeted PAHs. There was a weak positive correlation, but not statistically significant ($p > 0.05$), between the enhancement ratio of PAHs in different aqueous phases and the octanol–water partition coefficient (K_{ow}) (Figure 6). The coefficients of determination (r^2) from the regression analysis obtained for 5× OF-OSPW, salts with 5× OF-OSPW, OSPW_(in), and salts were 0.43, 0.50, 0.41, and 0.10, respectively (Figure 6). Except for dibenzothiophene and fluorene, PAHs with larger K_{ow} values showed greater rates of desorption and measured concentrations.

4. CONCLUSION

The results of this study demonstrated that dissolved organic compounds in OSPW alone or in mixture with salts can enhance the release of PAHs from a solid medium of PDMS

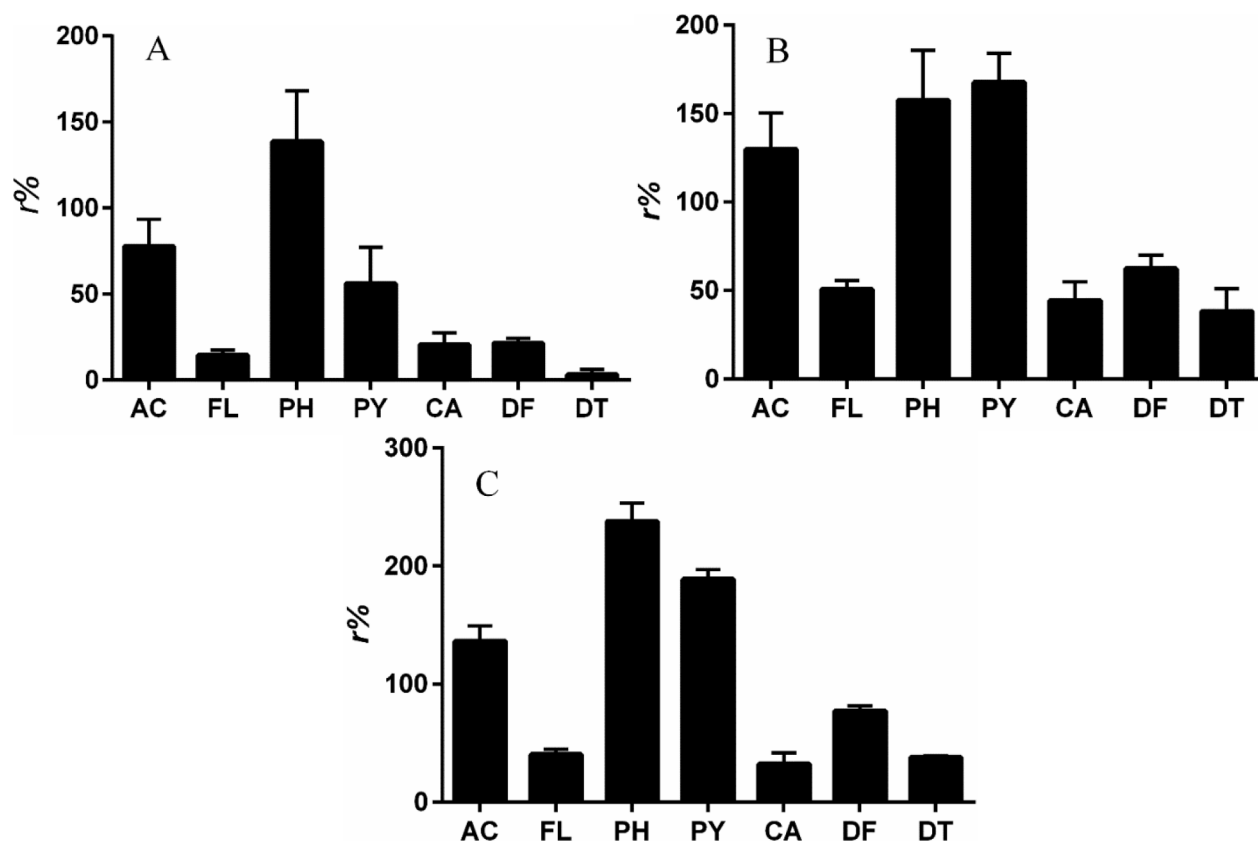


Figure 5. Effect of OF-OSPW on the increment ratio (r) of PAHs. The effects of (A) salts, (B) salts with 5 \times OF-OSPW, and (C) OSPW_(in) on the release of PAHs from PDMS to a water solution were measured once concentrations reached a steady state (see the calculation of r in Data Analysis).

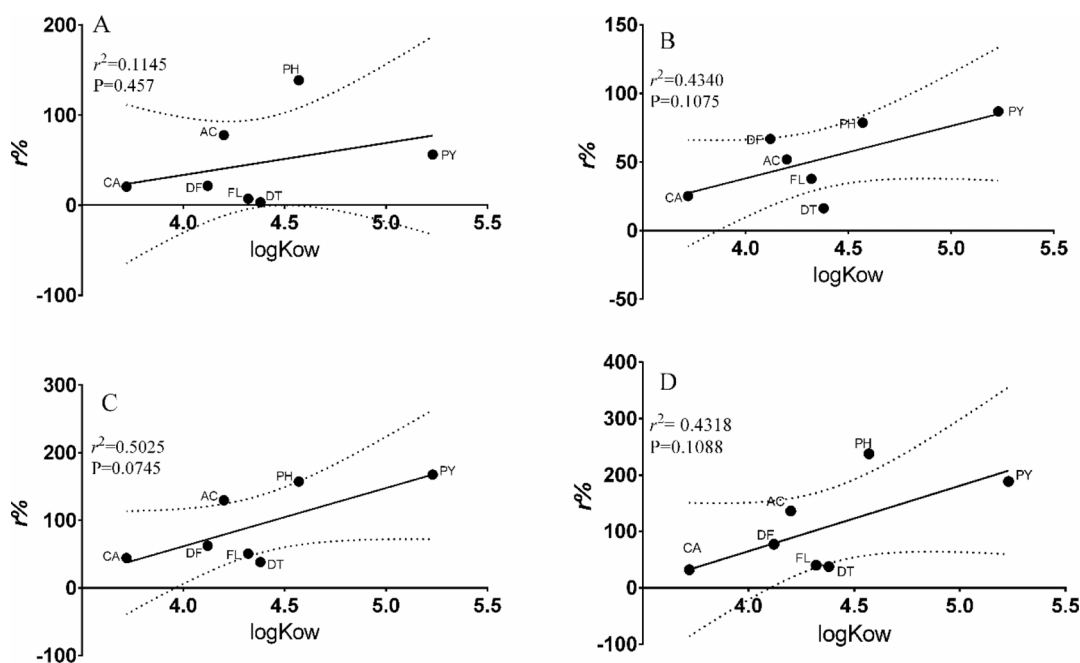


Figure 6. Regression analysis of (A) salts, (B) 5 \times OF-OSPW, (C) salts with 5 \times OF-OSPW, and (D) OSPW_(in) of the measured concentrations of PAHs loaded into PDMS, and $\log K_{ow}$. Values indicate the enhancement ratio (r) at 24 h of PAHs. The solid line indicates the regression, with associated correlation values displayed in each plot.

440 into the aqueous phase. This suggests that the concentrations
441 of PAHs in the aqueous phase of OSPW could increase as

PAHs partition from fine tailings. Future ecological risk
assessment and strategies for remediation of OSPW should 443

444 consider the effects of surfactants and salts on the
445 bioavailability of hydrophobic organic compounds, such as
446 PAHs, which could result in increased risks of the exposure of
447 aquatic organisms to PAHs.

448 ■ AUTHOR INFORMATION

449 Corresponding Author

450 **Hattan A. Alharbi** – Department of Plant Protection, College
451 of Food and Agriculture Sciences, King Saud University,
452 Riyadh 12372, Saudi Arabia; orcid.org/0000-0003-3297-729X;
453 Email: halharbii@ksu.edu.sa

454 Authors

455 **Steve B. Wiseman** – Department of Biological Sciences and
456 Water Institute for Sustainable Environments (WISE),
457 University of Lethbridge, Lethbridge, AB T1K 3M4,
458 Canada; orcid.org/0000-0002-8215-2272

459 **John P. Giesy** – Toxicology Centre and Department of
460 Veterinary Biomedical Sciences, University of Saskatchewan,
461 Saskatoon, SK S7N 5B3, Canada; Department of
462 Environmental Sciences, Baylor University, Waco, Texas
463 76706, United States; State Key Laboratory of Pollution
464 Control and Resource Reuse, School of the Environment,
465 Nanjing University, Nanjing 210023, People's Republic of
466 China

467 Complete contact information is available at:

468 <https://pubs.acs.org/10.1021/acsestwater.0c00183>

469 Notes

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