

Recent Developments in the Regulation of Monoamine Oxidase Form and Function: Is the Current Model Restricting Our Understanding of the Breadth of Contribution of Monoamine Oxidase to Brain [dys]Function?

Darrell D. Mousseau^{1,*} and Glen B. Baker²

¹Cell Signalling Laboratory, Department of Psychiatry, University of Saskatchewan, Saskatoon, Canada;

²Neurochemical Research Unit, Department of Psychiatry, University of Alberta, Edmonton, Canada

Abstract: Historically, much of the focus on monoamine oxidases and their substrates has been in the area of depression and the monoamine neurotransmitters serotonin (5-hydroxytryptamine), noradrenaline, and to a lesser extent, dopamine. With both forms of monoamine oxidase (A and B), the production of hydrogen peroxide as a byproduct of the reaction between the monoamine oxidases and their monoamine substrates has also implicated monoamine oxidase-sensitive events in intrinsic cell death pathways, particularly those centered on oxidative stress and peroxyradical-mediated mechanisms. Consequently, and perhaps not unexpectedly, the inhibition of monoamine oxidase has been considered as adjunctive therapy in neurodegenerative disorders such as Alzheimer's disease and Parkinson's disease, both of which involve a significant oxidative stress component. Yet the literature also provides ambiguities; indeed, not all of the functions of monoamine oxidases are dependent on catalytic activity nor can they all be ascribed to expression levels of the monoamine oxidase protein *per se*. Recent reports strongly suggest that the functions of monoamine oxidases also rely on post-translational modifications, epigenetic influences, interactions with other proteins, the cell phenotype and its localization to specific subcellular compartments. These recent developments certainly complicate the issue, yet they need to be duly considered when implicating monoamine oxidases and their inhibitors in both *in vitro* and *in vivo* pathological contexts.

Keywords: Monoamine oxidase; oxidative stress; apoptosis; phosphorylation; splice variant; catalytic independent; mitochondria; nucleus; Alzheimer disease; Parkinson's disease.

INTRODUCTORY COMMENTS

Neurodegenerative diseases are progressive diseases with symptoms that usually manifest following profound and irreversible cellular damage. In certain instances, such as with Huntington's disease, family history and genetics play an unequivocal role in the onset of the disease. In this particular disease the number of the trinucleotide 'CAG' repeats in exon1 of the *huntingtin* gene is inversely correlated with age of onset, i.e. the more repeats there are, the earlier and the more aggressive the disease [1, 2]. In other cases, the neurodegenerative diseases only emerge in the aged population, although there is debate as to whether the diseases, for example Alzheimer's Disease (AD), are 'age-related' or 'aging-related' [3]. The current review will examine the contribution of the enzyme, monoamine oxidase (MAO¹), to mental health and to neurodegenerative disease with a particular

emphasis on its role in the pathology of AD. We will provide some of the basic information required to familiarize the reader with MAO and we will examine some of the evidence, albeit oftentimes ambiguous, regarding the role(s) of MAO in neurodegeneration. We will occasionally include discussions of other pathologies in which MAO has been implicated to support or, in some cases, apparently disagree with our arguments. It is known that there are two isoforms of MAO, denoted MAO-A and MAO-B, with distinct potentials for contributions to neurodegeneration. Several recent reviews have examined the roles of MAO-B and some of its selective inhibitors, including dual/multi-target (e.g. MAO and acetylcholinesterase) inhibitors, in Parkinson's disease and in AD-related dementias [4-7]. In addition, an overview with a particular focus on the contribution of cell signalling and pro-/anti-apoptotic pathways in MAO-A inhibitor-mediated neuroprotection was recently published [8]. Rather than simply iterating the contents of these excellent reviews, the present review will comment on less obvious contributions by MAO-A, but the details will be contrasted and compared with MAO-B-related information when appropriate. While some of our comments might be provocative, they are certainly not intended to be critical. Our objective is to consider recent developments in the field, how these developments fit into the existing literature and, by extension, to provide novel perspectives on unintentional biases in the literature. We hope to highlight biases that might have steered the field away from a deeper understanding of MAO form and [patho]physiological function, and how these bi-

*Address correspondence to this author at the Cell Signalling Laboratory, GB41 HSB, University of Saskatchewan, 107 Wiggins Rd., Saskatoon, SK, S7N 5E5, Canada; Tel: +1-(306)-966-8824; E-mail: darrell.mousseau@usask.ca

¹The consensus for gene and protein nomenclature will be used throughout this review. Upper case letters will be used to designate the protein, regardless of species (e.g. MAO-A). Instances where upper case letters are italicized will refer to the human gene (for example *MAO-A*), whereas the mouse/rat gene will be italicized and will only have the first letter in upper case (for example, *Mao-A*). All mRNA and cDNA will follow the gene symbol formatting convention, for example *MAO-A* mRNA and *Mao-A* cDNA.

ases could have led to some of the intricacy and ambiguity in the associated literature.

THERE ARE TWO ISOFORMS OF MAO

MAO is a flavin adenine dinucleotide (FAD)-containing amine oxidase whose putative physiological function is the oxidative deamination of biogenic and xenobiotic monoamines. The reaction with MAO uses oxygen and results in the generation of the short-lived corresponding aldehyde as well as ammonia and hydrogen peroxide (H_2O_2) as reaction by-products. If these by-products are not detoxified, their accumulation is invariably toxic. For example, if H_2O_2 is not inactivated by glutathione peroxidase, then it can be converted by transition metal-mediated Fenton reactions to toxic hydroxyl radicals that can initiate lipid peroxidation and cell death [9]. This is exacerbated in situations where free radical scavenging or buffering systems may be compromised such as in the elderly [10] and/or during neurodegenerative processes [11]. There is strong evidence that oxidative stress plays a crucial role in the initiation and progression of AD [12, 13]. This is supported by the observation that reactive oxygen species, such as H_2O_2 , can mediate the neurotoxicity associated with the β -amyloid peptide [14, 15] and by the elevated oxidative damage in transgenic mouse models of AD [16, 17].

MAO, initially named tyramine oxidase given its ability to deaminate tyramine [18], was subsequently given the more inclusive designation, MAO, once the biogenic monoamines epinephrine, norepinephrine, serotonin (5-hydroxy tryptamine, 5-HT) and dopamine were also identified as substrates [19]. Two isoforms of MAO, i.e. MAO-A and MAO-B, have been identified according to differences in their specificities for inhibitors [20-22] and substrates [22-25]. MAO-A is inhibited by clorgyline at low nanomolar concentrations and this irreversible inhibitor has been used to estimate the turnover rate of MAO-A to be 2 days and recovery of MAO-A activity and protein levels after treatment with clorgyline to occur within 14 days [26]. MAO-B is inhibited by the irreversible inhibitor selegiline (l-deprenyl) at low concentrations, and studies using radiolabeled selegiline estimate the half-life of MAO-B in human brain to range between 30 and 40 days [27, 28]. Serotonin, noradrenaline, and adrenaline are preferential substrates for MAO-A, and benzylamine and β -phenylethylamine for MAO-B, yet there appears to be a functional mismatch between the specific isoform and its preferred substrate. Indeed, MAO-A (mRNA and protein) is highly expressed in catecholaminergic neurons of the locus coeruleus (and it is not found in serotonergic neurons), whereas MAO-B (mRNA and protein) is preferentially expressed in serotonergic neurons of the raphe nuclei, in histaminergic neurons and in glial cells [29-36]. This pattern of mismatch is conserved across species and is thought to be a means of mitigating off-target effects by amines diffusing in from adjacent regions/synapses [32, 33]. This notion is clearly illustrated by the fact that MAO-A is localized to noradrenergic nerve terminals of the rat pineal gland, while MAO-B is concentrated in juxtapositioned (serotonergic) pinealocytes [37]. Dopamine is a substrate for both isoforms in humans [38, 39] and preferentially for MAO-A in rats [23, 40]. In most species, dopamine, tyramine, and tryptamine are common substrates for both

MAO isoforms [22, 41, 42]. However, these substrate specificities are not absolute as both enzymes show broader substrate preference at high substrate concentrations [24]. This is an important consideration as MAO-B is known to play a central role in dopamine degradation in glia/astrocytes, yet MAO-A has also been detected in glia [43]. This, in combination with the therapeutic potential of non-selective MAO-A/B inhibitors [44], suggests that glial MAO-A could contribute to dopamine degradation when levels of this substrate are in excess [45].

The role of MAO-B in neurodegeneration has been widely studied and recently discussed [4-7, 46]. In contrast, the degradation of serotonin, noradrenaline and dopamine (and any associated cellular dysfunction) by MAO-A has been historically associated primarily with the neurobiology, and treatment, of depression [44, 47, 48] rather than any neurodegenerative phenotype. However, aspects of depression and progressive neurodegeneration could certainly rely on overlapping molecular mechanisms and could account for the recent spate of reports in the literature associating comorbid depression with many other disorders, including an increased risk of dementia and AD.

MAO-A and MAO-B are encoded by two different genes [49] located tail-to-tail on the Xp11.23-Xp 22.1 short arm [50, 51]. Both genes are comprised of 15 exons that span at least 60kb with an identical exon-intron organization that suggests duplication of a common ancestral gene [52]. Tissue-specific differences in the regulation of *MAO-B* transcription could rely on polymorphisms including a C-1,114T in the 5' region, a variable number 'GT' repeat in intron 2, and a G-to-A point mutation in intron 13 of the *MAO-B* gene [53]. A variable number tandem repeat (VNTR) polymorphism in the *MAO-A* promoter has five alleles containing 2, 3, 3.5, 4, or 5 copies of a 30-bp tandem repeat. Of these alleles, only those with three or four copies of the VNTR are common in different human populations, and those with 3.5 and 4 copies of the VNTR are transcribed much more efficiently than the alleles with 3 and 5 repeats [54]. These differences in transcription efficiency could account for the significant variability in MAO-A activity in different human skin fibroblast cultures [55] and could account for risk in Parkinson's disease [56], AD [57], impulsivity [58] and other neuropsychiatric disorders [59], although there is still debate as to the role of VNTRs in mood and depression [60]. It should be noted that imaging studies have clearly demonstrated that seemingly modest increases in MAO-A binding capacity (i.e. 34%) can account for the depressive phenotype in treatment-naïve depressed patients [61] as well as the depression associated with post-partum [62] and following smoking cessation [63]. It is interesting that prenatal exposure to cigarette smoke, which has long been known to contain an MAO inhibitory substance [64], is consistently associated with increased rates of behavioural problems, irritability, and attention-deficit/hyperactivity disorder [65], adolescent onset of drug dependence [66], and risk of violent offenses [67] and criminal arrest [68] in the offspring. MAO-B inhibition could clearly be contributing to the reduced incidence of Parkinson's disease in smokers [69] (with a possible predisposition to smoking by any one of the polymorphisms in the *MAO-B* gene already discussed [53, 70]). However, the neuropsychiatric and conduct disorders men-

tioned above would be more in keeping with a reduction in MAO-A availability as corroborated by recent neuroimaging studies [63, 71]. This long-lasting effect in the offspring strongly suggests an epigenetic modification, as does the much earlier finding of a region-dependent “daily rhythm of MAO” in human brain extracts [72]. An epigenetic component to the regulation of MAO function is now a strong consideration in the relevant literature. Indeed, hypermethylation of the *MAO-B* promoter has recently been linked to smoking [73] and epigenetic regulation of the *MAO-A* gene has been associated with several behavioural phenotypes [31, 74, 75] including a gender-specific propensity for nicotine dependence [76]. Tranylcypromine, a non-selective MAO inhibitor, is also a potent inhibitor of the histone demethylase, LSD1 [77], while the deacetylation (and activation) of the *MAO-A* promoter has been linked to angiogenic behaviour in the rat [31] and diurnal fluctuations in mood and MAO-A appear to be linked to components of the circadian clock [78]. These transient changes in MAO-A expression associated with epigenetic influences are not in accord with the earlier estimates of turnover rates and half-life for MAO-A that fall within the range of days to weeks [26]. Finally, it is vital to understand that the *MAO* gene could undergo tissue-specific splicing, as proposed for tissue differences in imidazoline binding [79] and as shown to occur following insertional mutagenic disruption of the *Mao-A* gene in mice [80]. We will re-visit the potential contributions by splice variants in our subsequent discussion.

RECENT OBSERVATIONS REVEAL AN EFFECT OF MAO-A THAT IS INDEPENDENT OF ITS CATALYTIC ACTIVITY

Exon 12 is the most conserved exon between *MAO-A* and *MAO-B* (and across species), ostensibly because of the importance of the functional domain that it encodes, i.e. a 33-amino acid expanse that contains the Ser-Gly-Gly-Cys-Tyr pentapeptide that flanks the cysteine406 residue (Cys406) to which the cofactor FAD covalently binds [81]. It is important to note that FAD is not an absolute requirement for full functionality of the MAO-A enzyme, but that it might play a substantive role in maintaining the structural integrity and stability of the enzyme [82, 83]. This seemingly minor observation has a significant impact on the interpretation of MAO-A function. Indeed, we have recently shown that overexpression of an MAO-A protein bearing the Asp328Gln substitution (already known to inhibit MAO-A activity [84]) was able to alter cell proliferation and *de novo* DNA synthesis in the human HEK293 cell line [85]. Furthermore, this same catalytic-dead variant was able to induce the expression of Bcl-2 and Bcl-XL, two anti-apoptotic/pro-survival molecules that also have been associated with pharmacological inhibition of MAO-A [86, 87]. Interestingly, we observed similar effects in breast cancer cells overexpressing a catalytic-dead Cys406-substituted MAO-A variant (Mousseau, Kuski, Pennington, unpublished data), which, as mentioned above, would mitigate FAD binding. These data strongly suggest that MAO-A-mediated events are not necessarily predicated solely on its catalytic activity. Until recently, the basic premise was that the catalytic activity of a given sample preparation was a valid reflection of the availability and expression of MAO-A protein within the sample. With the

commercial availability of relatively specific antibodies, it has become clear that this premise is flawed. Without this knowledge, the effect(s) of MAO have been attributed exclusively to catalytic reactions and not to any uncharacteristic catalytic-independent influences. This recent development certainly primes any debate on the exact role of MAO-A in physiological or pathological phenotypes.

Indeed, there is clear ambiguity in the relevant literature. For example, AD-related pathology is thought to rely primarily on the MAO-B isoform [88, 89] since MAO-B activity and *MAO-B* mRNA have been reported to be increased in platelets of AD patients [90, 91], as well as in the hippocampus, thalamus, and cerebral cortex, which are regions that undergo extensive neuronal cell death during AD [92, 93]. The MAO-B increase could be due to the local infiltration of glial cells in these areas as MAO-B is mainly expressed in this cell type, a significant proportion of which are found in the proximity of β -amyloid plaques (a hallmark of AD pathology) [32, 88, 94]. MAO-A activity and *MAO-A* mRNA are also reported to be elevated in several areas of the AD brain including the occipital cortex, frontal lobe of neocortex, parietal cortex, and locus coeruleus [93, 95, 96] as well as in the caudate nucleus, thalamus and white matter [97]. Although there are also reports of decreases in MAO-A activity in AD brains [95, 98], it is important to realize that a substantial amount of MAO-A (i.e. 75-80%) needs to be inhibited before any effects on MAO-A-mediated cell function would be notable [99, 100]. Furthermore, the 17-31% decrease in MAO-A activity in the AD locus coeruleus, where nearly 70-80% of the neurons are lost [95, 98, 101], suggests that the average MAO-A activity per *surviving* neuron is actually increased [95]. Such a localized hyperactivation of MAO-A could certainly account for the accumulation of toxic MAO-mediated metabolites in AD brains [102]. Similarly, in the earliest stages of AD, such a ‘hyperactivated’ form of MAO-A in MAO-A-immunoreactive cholinergic neurons in the nucleus basalis of Meynert and any H₂O₂-associated increase in oxidative stress could account for the excessive loss of cholinergic neurons in this structure [98] as well as in MAO-A-expressing serotonergic neurons in the dorsal raphe nucleus and noradrenergic neurons in the locus coeruleus [103-107]. We will re-visit the potential for a hyperactivated state of MAO-A in an AD-related context a little further on. For the moment, it is clear that estimating MAO-A activity without demonstrating corroborative changes in protein expression presents a significant bias in defining an MAO-A-mediated contribution to earlier stage disease progression based on extrapolations made using post-mortem, i.e. terminal stage, tissues (or any other model preparation, for that matter).

Estrogen, a neuroprotective hormone, can selectively decrease MAO-A activity and *Mao-A* mRNA levels in many brain areas [108, 109] and is thought to explain, in part, the increased risk of AD in estrogen-deficient, post-menopausal women. In serum withdrawal-induced neuronal apoptosis, MAO-A activity is selectively increased as is the activation of the pro-apoptotic enzyme caspase-3 [110, 111]. Parenthetically, impaired caspase-3/-9 expression following targeted siRNA-mediated MAO-A knockdown or R1-mediated repression of *Mao-A* transcription correlates with dysregulated apoptosis and disturbed neurodevelopment in an *in*

in vitro model of embryogenesis [112]. H₂O₂ generated by MAO induces cell apoptosis in kidney [113], while MAO-A, but not MAO-B, can bind with an endogenous neurotoxin to reduce mitochondrial membrane potential ($\Delta\psi_m$), thus providing additional mechanisms linking MAO-A to apoptotic cell death [114].

Yet, as with the discussion on AD above, the role of MAO-A in normal aging is equally ambiguous [96, 115-117] and if we examine the cancer literature, where MAO-A has recently been associated with disease progression, a similar ambiguity emerges. For example, *MAO-A/Mao-A* mRNA is decreased across all cancers (regardless of species) and this generalized decrease is proposed as a marker for tumour progression [118]. Yet our own studies reveal that the MCF-7 cell line has virtually no MAO-A activity, but the highly aggressive MDA-MB-231 breast cancer cell line has very high MAO-A activity (Satram-Maharaj, Nyarko, Kuski, and Mousseau, *unpublished data*). MAO-A activity is also increased in experimental breast cancer in rats [119]. Paradoxical increases in serotonin (which would contradict a putative concurrent increase in MAO-A activity) in human breast cancer are thought to support tumour growth [120]. MAO-A protein is also induced in high-grade prostate cancer [121], yet, again paradoxically, serotonin is concurrently increased [122] to the point that it (e.g. serotonin) is also proposed as a valid marker for prostate tumour progression [123]. It is interesting that these reports base their conclusions on MAO-A protein expression and do not include any estimate of MAO-A activity. Without any evidence to the contrary, the MAO-A protein detected in these reports could be in an *inactive* form and, as our work with the MAO-A(Asp328Gln) catalytic-*dead* variant shows [85], could be promoting proliferative phenotypes *via* non-catalytic-based *de novo* DNA synthesis and/or induction of anti-apoptotic Bcl-2-related proteins.

Could a pool of catalytic-*dead* MAO-A protein influence brain function? While this has never been pointedly examined, there is evidence, albeit indirect, in support of this possibility. For example, our recent work demonstrates that mice expressing the AD-related M146V-substituted presenilin-1 (PS-1) protein express significantly more cortical MAO-A protein than their wildtype littermates [124]; however, MAO-A activity in these mice remains comparable to wildtype levels, which suggests that the *de novo* pool of MAO-A protein is somehow rendered *inactive*. In these PS-1(M146V) mice there is a significant disruption of cortical cytoarchitecture and laminar organization [124] that is a phenotype reminiscent of the disrupted cortical lamination observed in both prenatal PS-1-null mice [125] and postnatal PS-1 conditional knockout mice [126]. This certainly implicates the PS-1 variant itself in this phenotype. Yet, similar permanent cytoarchitectural alterations are also evident in the somatosensory cortex of MAO-A-deficient mice [80], which, presumably, do not bear a PS-1 defect. One could argue that it is the hyperserotonergic tone that is observed in both PS-1(M146V) [124] and MAO-A-deficient [80] mice that is the commonality underlying the cytoarchitectural alteration in these different strains of mice. This, again, is a reasonable assumption given the similar cortical disruptions associated with hyperserotonergic (but not noradrenergic) tone produced by administration of the MAO-A inhibitor

clorgyline to normal mice during their first week of life [127] or to the clorgyline-induced neurodevelopmental changes observed during *in vitro* embryogenesis [112]. Yet one could also argue that it is MAO-A in its *inactive* state that is also partly responsible for these effects. While such a pool is present in the PS-1(M146V) mice [124] and an inactive human MAO-A protein does exert phenotypic changes *in vitro* [85], how would this mechanistically relate to the original *Mao-A*-deficient mouse generated by Cases and colleagues [80]? In fact, this is where it is important to recall that these mice were generated by insertional mutagenesis (i.e. the gene for interferon- β was inadvertently inserted antisense into exon2 –and thus disrupting normal transcription of the *Mao-A* gene); the authors clearly demonstrated that the normal, i.e. full length, *Mao-A* transcript was absent, but that there was evidence of exon skipping, resulting in four detectable *Mao-A* splice variants. This raises the distinct possibility that ‘exon-deleted’ MAO-A proteins exist and while any MAO-A protein expressed in these mice would clearly be devoid of catalytic activity (not surprising, given the loss of exon2 and the portion of the FAD-binding domain encoded by this exon), the existence of inactive MAO-A protein variant(s) based on the remaining exons could certainly be contributing to various phenotypes. Finally, one must extend these suppositions to the ‘knock-out’ mouse used by the Shih laboratory and generated as a result of a spontaneous A863T point mutation in the *Mao-A* gene coding sequence that results in a ‘TAA’ codon and a premature ‘stop’ during the translation of the MAO-A protein [128]. There is a complete loss of MAO-A catalytic activity in these mice, as would be expected of the truncated gene product, but the possibility of exon skipping has never been examined (or excluded). Thus, the possibility that a deletion mutant MAO-A protein could be triggering such profound developmental changes in these two ‘knock-out’ mice and potentially exerting some influence on the proliferation of neural stem cells, as observed recently in *Mao-A/B* ‘knockout’ mice [129], is certainly intriguing and warrants further investigation. Could similar splice variants be contributing to the behavioural and cognitive phenotypes in the human kindred bearing a point mutation in exon8 of the *MAO-A* gene and functional deletion of the MAO-A protein [130]? This is also worthy of consideration.

Finally, one has to wonder whether a catalytic-*dead* MAO-A protein could affect other neurodegenerative diseases with a MAO-based causative mechanism. For example, Parkinson’s disease has been historically associated with an MAO-B-sensitive etiology as well as therapies based on the putative MAO-B-selective inhibitors selegiline and rasagiline [5]. Rasagiline can also prevent the caspase activation and oxidative stress associated with an *in vitro* α -synuclein model of parkinsonism [131]. Recent work, however, has suggested that not all of the pathology in rodent models of Parkinson’s disease can be ascribed solely to MAO-B [132]. Indeed, some of the neuroprotective effects of rasagiline, particularly its ability to induce anti-apoptotic proteins such as Bcl-2-related proteins [133], might reflect MAO-A-mediated mechanisms in human SH-Sy5y neuroblastoma cells [8, 87]. In fact, there is a reasonable evidence that the N-propargyl moiety (found in many MAO inhibitors, e.g. pargyline, clorgyline, selegiline, rasagiline, ladostigil)

can exert effects and activate cellular signalling cascades (particularly those associated with neuroprotection/rescue) independent of any catalytic inhibition of the targeted MAO enzyme [5, 44, 134, 135]. Although it remains to be determined whether non-catalytic properties of an off-target inhibition of MAO-A might be responsible for any Parkinson's disease-modifying effects of rasagiline or selegiline, our work on the catalytic-dead MAO-A(Asp 328Gln) variant and its ability to induce Bcl-2-related proteins [85] certainly support this possibility.

FIRST-GENERATION (IRREVERSIBLE) VERSUS NEWER REVERSIBLE INHIBITORS OF MAO

The substrate binding site is similar in MAO-A and MAO-B, suggesting that substrate and inhibitor specificities rely on additional influences, including the size of the recognition site itself, which is smaller in MAO-B [20]. The region contained within residues 120-220 and residues 50-400 determines substrate preference of rat liver MAO-A and MAO-B, respectively, while the region flanked by residues 220-400 appears to contribute to the relative catalytic activity towards their respective substrates [136]. Furthermore, crystal structures of the cavity-shaping loop at residues 210-216 in human MAO-A and 201-206 in human MAO-B [137] implicate these regions in substrate recognition, while residues 89-219 and 295-399 of human MAO-A may contribute to substrate/inhibitor-binding domains [137]. This is supported by the loss of catalytic activity associated with the deletion of carboxy-terminal amino acids in human MAO-A [138, 139] and explains why substitution of the carboxy-terminal amino acids of MAO-B with those of MAO-A imparts MAO-A activity and inhibitor specificity to the chimeric MAO-B/A protein [140].

The mechanism-based inhibition by irreversible 'suicide' inhibitors, such as clorgyline or selegiline, occurs following their binding to the FAD cofactor [141]. This binding triggers MAO to process clorgyline and selegiline as if they were substrates. As the catalytic reaction proceeds, a reactive intermediate covalently alkylates FAD, which effectively and irreversibly (hence 'suicide') blocks subsequent access by substrate(s). The so-called 'cheese effect' is a well-documented, noxious side-effect of irreversible inhibition of gut wall MAO-A, the resulting in the elevation in circulating levels of dietary sympathomimetic amines such as tyramine and, ultimately, tyramine-induced release of noradrenaline. Symptoms that can range from headache to a hypertensive crisis were first associated with MAO inhibitors and high tyramine content foods such as certain red wines and aged cheeses (hence 'cheese effect'). Concern about hypertensive crises led to the development of reversible inhibitors of MAO-A (RIMAs). Because tyramine still has competitive access to the active site on MAO-A and the 'cheese effect' is thus avoided, RIMAs such as moclobemide are better tolerated by at-risk populations, particularly the elderly, including those with cognitive deficits [142, 143]. RIMAs have a further advantage over irreversible inhibitors in that there is usually full recovery of brain MAO within 24 hours after cessation of treatment [144], which is an important consideration if transferring a patient onto a drug regimen that may be contraindicated with elevated levels of biogenic amines.

MAO-A expression could be a risk factor for AD [57, 93, 145, 146]. Depression, perhaps by virtue of its capacity to promote cognitive impairment, is now thought to represent a prodrome for AD-related dementia in certain patients [147-149]. This suggests that MAO inhibitors could be useful adjunctive drug therapies in AD. RIMAs are particularly efficacious in treating depression [142] and cognitive disorders [143] in the elderly, and are potential anti-apoptotic agents [150]. The selective MAO-A inhibitor clorgyline inhibits glutamate-induced excitotoxicity [151] as well as apoptosis induced by serum starvation [110, 150] and by the AD-related β -amyloid peptide *in vitro* [15], and protects against damage caused by the mitochondrial toxin malonate *in vivo* [152]. While selegiline is often used in Parkinson's disease [5], its benefit in AD patients remains a matter of debate [153, 154]. However, dual inhibitor drugs, i.e. those that target MAO and acetylcholinesterase simultaneously, have been proposed as therapeutics in AD [155], and ladostigil, which combines the activity of rasagiline and anticholinesterase activity, provides some benefit in the context of AD [156].

Depression is also a prodrome in Parkinson's disease [157] and appears to reflect reductions of noradrenaline in the locus coeruleus and of serotonin in the raphé nucleus [158]. While the combination of the older, irreversible MAO inhibitors, e.g. clorgyline and selegiline, with the Parkinson's therapeutic dopamine precursor, l-dopa, had raised some clinical concern [159, 160], the same does not hold for the newer generation MAO inhibitors. It is clear that the RIMAs moclobemide [161], brofaromine [162], and bexloxtone [163], because they can be competitively displaced from the MAO-A enzyme by excess dopamine, also allow any excess dopamine (and other biogenic amine substrates) to be degraded [164]. Thus, Parkinson's patients tolerate moclobemide because of a limited 'cheese effect' [165]; because it improves motor functions [166]; and because it benefits the significant proportion of this patient population who suffer from depression [158, 167].

DO THE NEUROPROTECTIVE EFFECTS OF MAO INHIBITORS RELY SOLELY ON CATALYTIC INACTIVATION OF THE ENZYME?

While it is assumed that MAO inhibitors are usually quite selective and specific, in fact there are many reports that MAO inhibitors interact with other amine oxidases, various transaminases, decarboxylases, dehydrogenases, cytochromes, cytochrome P450s, biogenic amine receptors and transporters, imidazoline binding sites, and even sigma receptors (see reviews [168, 169]). All of these off-target interactions could certainly contribute to the therapeutic and/or adverse effect profiles of MAO inhibitors. Interestingly, the selective serotonin reuptake inhibitor (SSRI) antidepressants fluoxetine and its active metabolite nor-fluoxetine have been shown to have MAO-A inhibitory properties in the rat [170] and preliminary investigations (Mousseau, Holt and Baker, *unpublished data*) indicate that the same may hold true for human MAO-A, which could be part of the reason that the use of these SSRIs is contraindicated in patients already on an MAO inhibitor regimen.

Selegiline and rasagiline have been reported to prevent apoptotic phenotypes by up-regulating the anti-apoptotic

Bcl-2 and by down-regulating the pro-apoptotic Bad and Bax, and preventing nuclear translocation of glyceraldehyde-3-phosphate dehydrogenase [44]. Selegiline also has been reported to inhibit the accumulation and fibrillar behaviour of β -amyloid [171] and can reverse age-related memory impairment [172]. Furthermore, pretreatment with selegiline or pargyline (also an irreversible inhibitor of MAO-B) protects dopaminergic neurons against MPTP-mediated neurotoxicity *in vivo* [173, 174] and *Mao-B* knock-out mice are resistant to MPTP-induced neurotoxicity [35]. Yet the effects of MAO-B inhibitors in some of the examples mentioned above might not rely specifically on MAO-B itself since neuroprotection was often associated with concentrations of the drug that were too low to inhibit the enzyme [175, 176]. Phenelzine, an irreversible inhibitor of MAO-A and MAO-B, also elevates brain levels of GABA, alanine and ornithine, sequesters toxic aldehydes such as 3-aminopropanal, acrolein and formaldehyde, and inhibits the enzyme primary amine oxidase [168, 177-180]; some of these actions could well be contributing to its reported neuroprotective effects [181]. Both tranylcypromine (also an irreversible inhibitor of MAO-A and MAO-B) [182] and phenelzine [183] have been shown to induce the expression of brain-derived neurotrophic factor in rat brain. Furthermore, the (S)-isomer of rasagiline (TVP1022), which possesses the N-propargyl moiety, but does not inhibit MAO-B, is neuroprotective [184], whereas rasagiline can induce both GDNF mRNA and protein expression, but an analogue devoid of the N-propargyl moiety cannot [185]. This suggests actions of these MAO inhibitors that could rely on an innate action of this moiety (as we had already discussed above), rather than on any catalytic-dependent mechanism. In addition, N-propargyl-containing compounds, such as selegiline and rasagiline, have been found to activate Bcl-2 family members, elevate superoxide dismutase and glutathione levels, up-regulate tyrosine hydroxylase and aromatic amino acid decarboxylase [134, 135, 186, 187], and interact with the mitochondrial pore complex and modulate amyloid precursor protein cleavage [44, 188, 189]. More recently, tranylcypromine has been shown to be a potent inhibitor of the histone demethylase, LSD1 [77], which implicates epigenetic regulation in its range of mechanism of action(s). Any of these mechanisms of action would certainly provide for elements of neuroprotection.

The use of the clorgyline in our recent characterization of the physical interaction between the AD-related PS-1 (M146V) protein and MAO-A revealed a surprising observation. It is known that AD-related mutations can exert changes in the PS-1 protein that, in turn, influence the conformation of the PS-1-substrate complex [190]. Perhaps by virtue of the direct interaction between PS-1(M146V) and MAO-A, a concomitant change in the structure of MAO-A could be occurring that could account for the increased potency of clorgyline we observed in the PS-1(M146V) mouse brain samples. Specific residues in MAO-A have been associated with conformational stability and access to the catalytic cleft [191]. It is therefore not unreasonable to posit that a PS-1-induced conformational change in the MAO-A protein could alter the accessibility of clorgyline to its binding site and would suggest partially interconvertible states of a single clorgyline binding site. Yet, perhaps more impor-

tantly, the increased potency of clorgyline, as a mechanism-based inhibitor, implies that the MAO-A protein in the PS-1(M146V) brain is in actual fact far more active, but is presumably being maintained in a latent, *inactive* state by the PS-1 protein. If cellular events could disrupt the physical interaction between PS-1 and this hyperactive MAO-A, then could a localized surge in MAO-A/H₂O₂-mediated oxidative stress ensue that could overwhelm free radical-scavenging coping mechanisms? Such a scenario could certainly contribute to the average increase in MAO-A activity per *surviving* neuron in vulnerable AD brain regions [95] and, as already suggested above, could certainly contribute to both the accumulation of toxic MAO-mediated metabolites in AD brains [102] as well as the loss of MAO-expressing neurons in the dorsal raphe nucleus, the locus coeruleus and the nucleus basalis of Meynert [98, 103-107].

MOLECULAR BIOLOGY HAS IDENTIFIED AMINO ACID RESIDUES AND DOMAINS THAT ARE CRITICAL FOR MAO FUNCTION

The study of the three-dimensional structure of MAO has advanced due, in large part, to investigations using selective irreversible inhibitors [192]. There is much similarity between human MAO-A and MAO-B, although a major difference is that human MAO-B is dimeric, whereas human MAO-A crystallizes as a monomer [193]. Also, the cavity-shaping loop is larger in human MAO-A than in human MAO-B or rat MAO-A (*note*: rat MAO-A crystallizes as a homodimer), suggesting that this cavity-shaping loop is involved in the process of dimerization [137]. A single amino acid residue, e.g. Ile335 in MAO-A and Tyr326 in MAO-B, dictates substrate specificity and sensitivity to selective inhibitors in the corresponding enzymes [84, 194]. Phe208 in rat MAO-A and Ile199 in rat MAO-B are also reported to contribute to substrate and inhibitor specificities [195], although mutations of these two corresponding residues in human MAOs do not alter substrate specificity [196]. Ser209 in human MAO-A also contributes to MAO-A function, but the same does not hold true for the analogous residue in MAO-B (i.e. Ser200) [197]. Other studies have identified specific lysine, tryptophan and tyrosine residues [198] and cysteine residues [199, 200] that may contribute either to FAD binding or to stabilizing the protein's conformation, access to the catalytic cleft and its substrate binding capacity [191, 198].

NON-MITOCHONDRIAL LOCALIZATIONS OF MAO PROTEINS HAVE BEEN OVERLOOKED

Recently, crystallography has revealed that the carboxy-terminal amino acids 463-506 in human MAO-A are responsible for membrane anchoring [137]. Studies on the membrane insertion region in rat liver MAO-B reveal that deletion of the 28 carboxy-terminal amino acids blocks the localization of MAO-B to mitochondria. The cytochrome b5 protein fused with the carboxy-terminal 28 amino acids of rat MAO-B is found expressed in mitochondria instead of remaining in cytoplasm [201], suggesting that the mitochondrial targeting signal of rat MAO-B is located within this region [201]. The identical region in human MAO-B appears to determine mitochondrial localization [138, 202]. It is quite

possible that these regions contain the ubiquitination site necessary for insertion of both MAO-A and MAO-B into the mitochondrial membrane [203, 204].

It is interesting that a catalytically active conformation of MAO-A is detected immediately upon interacting with the mitochondrial outer membranes, but prior to its ubiquitin-dependent insertion into the membrane [203], which suggests transient, but distinct pools of MAO-A and/or a conformational reconfiguration of the MAO-A by virtue of an interaction with a molecule already present on the outer membrane of the mitochondrion. The notion of pools of MAO-A is not novel; indeed, mitochondria are thought to contain approximately 70% of the total cellular MAO activity, whereas the microsomal fraction accounts for approximately 25% and the balance is thought to be present in a 'soluble' form [205, 206]. This distribution is not a consistent, however, as the heart seems to have a disproportionately high level of microsomal and 'soluble' MAO relative to that detected in mitochondria [207].

It should be noted that MAO activity in the lysosomal fraction was significantly lower under control conditions than when rats were treated with ^{14}C -pargyline, which indicated that the lysosomes rapidly accumulated labeled MAO [208]. These authors also noted that the rate of return of MAO was much more rapid in the microsomal fraction than in the corresponding mitochondrial fraction, suggesting that the lysosomal pool of MAO could be a 'precursor' for the mitochondrial pool, although this was never firmly established. These same authors observed that the yields of protein obtained in various fractions were not similar to the recoveries of enzyme activity or radioactivity. In retrospect, this mismatch of MAO protein and catalytic activity was perhaps the first indication of a possible post-translational regulation of MAO function.

An immunohistochemical study clearly showed that pargyline-sensitive pools of MAO could be detected in several subcellular compartments, including the rough endoplasmic reticular membranes, mitochondrial outer membranes, within the nuclear envelope and along parts of the plasma membranes in diverse tissues [209]. MAO activity has also been associated earlier with the nuclear membrane [210] and more recently the MAO-A protein, although having significantly lower catalytic activity, was found to re-locate to the nuclear fraction during pre-eclampsia/eclampsia [211]. The reduction in MAO-A activity in pre-eclampsia was confirmed, but the relocalization to the nucleus was not [212]. Pools of synaptic and extrasynaptic MAO have also been confirmed and, according to the authors, studies using total homogenates could provide misleading information because a substantial reduction of activity within a specific cellular location (e.g. in the synaptic terminals) could be masked by measurement of the total activity in tissue homogenates [213]. Although the evidence for expression of MAO proteins in diverse subcellular compartments exists, this simple, yet critical, fact is often overlooked in the interpretation of results. Indeed, MAO is now conveniently referred to as the 'mitochondrial enzyme' and this is such a commonly accepted fact that MAO is often used as a marker of mitochondrial fraction purity. While interpreting results based on a purely 'mitochondrial' localization is 'convenient', it certainly biases our

understanding of the true role, and localization, of MAO in normal and pathological cell function.

POST-TRANSLATIONAL REGULATION OF MAO-A FUNCTION

This commentary would not be complete without an examination of post-translational regulation of MAO-A in disease. The induction of *MAO-A* mRNA and MAO-A protein and activity are known to correlate in the human encephalopathic brain [214]. This is not necessarily the case for the normal aging human brain [215] or for the AD brain, as mentioned above, and a discrepancy between *MAO* transcript levels in certain human and rodent cell lines has also been observed [216]. MAO-A, but not MAO-B, responds to manipulation of calcium (Ca^{2+}) either directly in rat brain homogenates [217] or by *in vivo* treatment with the Ca^{2+} -channel blocker nimodipine [218]. It is also known that Ca^{2+} selectively enhances MAO-A activity in mouse hippocampal HT-22 cells [15] and that the bacterial-derived toxin staurosporine can induce MAO-A-sensitive apoptosis in human neuronal SH-Sy5y cells [219], and that both of these latter effects occur independently of any change in *MAO-A* mRNA. This suggests the potential for post-translational regulation of MAO-A function. Although activation of the p38(MAPK) (p38 mitogen activated protein kinase) pathway has been associated with the induction of *Mao-A* mRNA and an MAO-A-sensitive apoptotic phenotype in PC12 and SK-N-BE(2)-C cells [110, 220], other studies [111, 216] clearly demonstrated that the effects of the p38(MAPK) protein itself on MAO-A-mediated events occur independently of changes in *MAO-A/Mao-A* transcript levels, respectively. We have identified Ser209 in overexpressed human MAO-A as a possible phosphorylation target for p38(MAPK) [197], although studies based on human MAO-A protein overexpressed in *Pichia pastoris* were not as conclusive [221]. It should be noted that human MAO-B has a homologous Serine residue (e.g. Ser200), but its cavity shaping loop is in a more compact conformation and there are no adjacent anionic groups near the hydroxyl of the Ser200 side chain to elicit a conformational change were it to be phosphorylated. These structural considerations might explain why p38(MAPK) could regulate MAO-A, but not MAO-B, *in vitro*. It remains to be seen if p38(MAPK) can be associated with the regulation of MAO proteins in human tissue preparations.

While any inhibitory effect of p38(MAPK) on MAO-A function would initially appear quite paradoxical, high basal activity of p38(MAPK) in healthy adult mouse brain has been observed, which suggests that this signalling protein, which is more often associated with a pro-apoptotic phenotype, might also contribute to normal brain cell physiology and survival [222, 223]. A pro-survival adaptive response to transient stress is suggested by the fact that H_2O_2 , the by-product of MAO reactions, can actually activate p38(MAPK) [224] and inhibit MAO-A activity [225], apparently *via* a Ca^{2+} -dependent mechanism [226]. The PI3K/Akt pro-survival pathway might also be involved in regulating aspects of MAO-A function, but its effect does not include any influence on the Ca^{2+} -mediated regulation of MAO-A function [216]. Valproic acid was found to induce *MAO-A* promoter function as well as catalytic activity *via* an Akt-FoxO1

(transcription)-sensitive mechanism [227]. In addition, the Jun N-terminal kinase [228, 229], Ras/ERK (p44/42) [216, 230] and TGF- β /Smad3 [231] signalling cascades have been implicated in the regulation of MAO expression, function and/or inhibition. Given the acknowledged role of these cascades in regulating both transient (post-translational) and long-term (genetic/epigenetic) effects of cell signalling and responses to extracellular cues, a closer examination of the influence of signalling pathways on the regulation of the MAO-A and MAO-B systems is clearly needed.

As we draw this review to a close we acknowledge that our discussion on the therapeutic targeting of MAOs has focused primarily on pharmaceutical inhibitors. It is clear that there is a broad range of naturally occurring regulators of MAO function that have been omitted herein, including, but clearly not limited to, caffeine and analogues [232], kaempferol [233], and dietary components and associated metabolites [234, 235]. We do apologize *a priori* for not having expanded upon this very important topic.

CONCLUDING REMARKS

MAO is often described as an ubiquitous, membrane-bound enzyme that is expressed on the outer membrane of the mitochondria. Our current understanding of the contribution of MAO proteins to neurodegenerative processes is very often centered on the production of hydrogen peroxide as a byproduct of the reaction between MAO and biogenic substrates. The interpretation of results invariably attempts to satisfy a convenient model that is predicated on the assumptions (a) that MAO is a passive enzyme (i.e. it simply waits for a substrate to metabolize), (b) that only the expression of MAO protein dictates catalytic activity in a given tissue, and (c) that MAO proteins are *only* expressed in the outer mitochondrial membrane. Given that MAO is anchored to the membrane through a very short C-terminal trans-membrane helix, much of the protein is presumably left exposed for potential modification(s) by, and/or interactions with, other proteins (either circulating or juxtaposed on adjacent membranes). Fluctuations in MAO function might reflect protein levels that are influenced by both polymorphisms and epigenetic influences. Furthermore, the potential for effects of MAO that are not solely dependent on its catalytic activity might be a strategy that has evolved to satisfy different functional requirements for the MAO enzymes in a context- or cell-dependent manner. In other words, it is possible that MAO proteins display a behaviour that lies between catalytically active and inactive states, and that at any given time a cell's phenotype would be influenced by the dominating state. Thus, simply using an estimate of MAO activity as an outcome, without a concurrent evaluation of MAO expression levels and/or its localization within different cellular compartments, could bias the interpretation of the experimental outcomes. Although this might initially be viewed as an unnecessary increase in workload, in the long-term this would provide for more information and would surely help to clear up some, if not a substantial portion, of the ambiguity that surrounds the role(s) of MAO proteins in [patho]physiological contexts.

CONFLICT OF INTEREST

The author(s) confirm that this article content has no conflicts of interest.

ACKNOWLEDGEMENTS

GBB holds a University of Alberta Distinguished University Professorship and funding from the Canadian Institutes of Health Research (CIHR). DDM holds the Saskatchewan Research Chair in *Alzheimer's Disease and Related Dementias* funded jointly by the Alzheimer Society of Saskatchewan and the Saskatchewan Health Research Foundation. DDM also holds funding from the Canadian Breast Cancer Foundation-Prairies/NWT Region. There are no competing interests.

REFERENCES

- [1] Schulte, J.; Littleton, J.T. The biological function of the Huntingtin protein and its relevance to Huntington's Disease pathology. *Curr. Trends Neurol.*, **2011**, *5*, 65-78.
- [2] Andrew, S. E.; Goldberg, Y. P.; Kremer, B.; Telenius, H.; Theilmann, J.; Adam, S.; Starr, E.; Squitieri, F.; Lin, B.; Kalchman, M. A.; *et al.* The relationship between trinucleotide (CAG) repeat length and clinical features of Huntington's disease. *Nat. Genet.*, **1993**, *4*(4), 398-403.
- [3] Ritchie, K.; Kildea, D. Is senile dementia "age-related" or "ageing-related"?--evidence from meta-analysis of dementia prevalence in the oldest old. *Lancet*, **1995**, *346*(8980), 931-934.
- [4] Schapira, A. H. Monoamine oxidase B inhibitors for the treatment of Parkinson's disease: a review of symptomatic and potential disease-modifying effects. *CNS Drugs*, **2011**, *25*(12), 1061-1071.
- [5] Riederer, P.; Laux, G. MAO-inhibitors in Parkinson's Disease. *Exp Neurol*, **2011**, *20*(1), 1-17.
- [6] Pisani, L.; Catto, M.; Leonetti, F.; Nicolotti, O.; Stefanachi, A.; Campagna, F.; Carotti, A. Targeting monoamine oxidases with multipotent ligands: an emerging strategy in the search of new drugs against neurodegenerative diseases. *Curr. Med. Chem.*, **2011**, *18*(30), 4568-4587.
- [7] Naoi, M.; Maruyama, W. Monoamine oxidase inhibitors as neuroprotective agents in age-dependent neurodegenerative disorders. *Curr. Pharm. Des.*, **2010**, *16*(25), 2799-2817.
- [8] Naoi, M.; Maruyama, W.; Inaba-Hasegawa, K.; Akao, Y. Type A monoamine oxidase regulates life and death of neurons in neurodegeneration and neuroprotection. *Int. Rev. Neurobiol.*, **2011**, *100*, 85-106.
- [9] Jenner, P.; Olanow, C. W. Oxidative stress and the pathogenesis of Parkinson's disease. *Neurology*, **1996**, *47* (6 Suppl 3), S161-170.
- [10] Lang, C. A.; Naryshkin, S.; Schneider, D. L.; Mills, B. J.; Lindeman, R. D. Low blood glutathione levels in healthy aging adults. *J. Lab. Clin. Med.*, **1992**, *120*(5), 720-725.
- [11] Schulz, J. B.; Lindenau, J.; Seyfried, J.; Dichgans, J. Glutathione, oxidative stress and neurodegeneration. *Eur. J. Biochem.*, **2000**, *267*(16), 4904-4911.
- [12] Christen, Y. Oxidative stress and Alzheimer disease. *Am. J. Clin. Nutr.*, **2000**, *71*(2), 621S-629S.
- [13] Gerlach, M.; Riederer, P.; Youdim, M. B. Molecular mechanisms for neurodegeneration. Synergism between reactive oxygen species, calcium, and excitotoxic amino acids. *Adv. Neurol.*, **1996**, *69*, 177-194.
- [14] Opazo, C.; Huang, X.; Cherny, R. A.; Moir, R. D.; Roher, A. E.; White, A. R.; Cappai, R.; Masters, C. L.; Tanzi, R. E.; Inestrosa, N. C.; Bush, A. I. Metalloenzyme-like activity of Alzheimer's disease β -amyloid. Cu-dependent catalytic conversion of dopamine, cholesterol, and biological reducing agents to neurotoxic H₂O₂(2). *J. Biol. Chem.*, **2002**, *277*(43), 40302-40308.
- [15] Cao, X.; Wei, Z.; Gabriel, G. G.; Li, X.; Mousseau, D. D. Calcium-sensitive regulation of monoamine oxidase-A contributes to the production of peroxyradicals in hippocampal cultures: implications for Alzheimer disease-related pathology. *BMC Neurosci.*, **2007**, *8*, 73.

- [16] Smith, M. A.; Hirai, K.; Hsiao, K.; Pappolla, M. A.; Harris, P. L.; Siedlak, S. L.; Tabaton, M.; Perry, G. Amyloid- β deposition in Alzheimer transgenic mice is associated with oxidative stress. *J. Neurochem.*, **1998**, *70*(5), 2212-2215.
- [17] Butterfield, D.A.; Castegna, A.; Lauderback, C.M.; Drake, J. Evidence that amyloid β -peptide-induced lipid peroxidation and its sequelae in Alzheimer's disease brain contribute to neuronal death. *Neurobiol. Aging*, **2002**, *23* (5), 655-664.
- [18] Hare, M. L. Tyramine oxidase: A new enzyme system in liver. *Biochem. J.*, **1928**, *22*(4), 968-979.
- [19] Blaschko, H.; Richter, D.; Schlossmann, H. The oxidation of adrenaline and other amines. *Biochem. J.*, **1937**, *31*(12), 2187-2196.
- [20] Finberg, J. P.; Youdim, M. B. Selective MAO A and B inhibitors: their mechanism of action and pharmacology. *Neuropharmacology*, **1983**, *22*(3 Spec No), 441-446.
- [21] Knoll, J.; Magyar, K. Some puzzling pharmacological effects of monoamine oxidase inhibitors. *Adv. Biochem. Psychopharmacol.*, **1972**, *5*, 393-408.
- [22] White, H. L.; Glassman, A. T. Multiple binding sites of human brain and liver monoamine oxidase: substrate specificities, selective inhibitions, and attempts to separate enzyme forms. *J. Neurochem.*, **1977**, *29*(6), 987-997.
- [23] Johnston, J. P. Some observations upon a new inhibitor of monoamine oxidase in brain tissue. *Biochem. Pharmacol.*, **1968**, *17*(7), 1285-1297.
- [24] Ochiai, Y.; Itoh, K.; Sakurai, E.; Adachi, M.; Tanaka, Y. Substrate selectivity of monoamine oxidase A, monoamine oxidase B, diamine oxidase, and semicarbazide-sensitive amine oxidase in COS-1 expression systems. *Biol. Pharm. Bull.*, **2006**, *29*(12), 2362-2366.
- [25] Fowler, C. J.; Callingham, B. A. Substrate-selective activation of rat liver mitochondrial monoamine oxidase by oxygen. *Biochem. Pharmacol.*, **1978**, *27*(16), 1995-2000.
- [26] Egashira, T.; Yamanaka, Y. Further studies on the synthesis of A-form monoamine oxidase. *Jpn. J. Pharmacol.*, **1981**, *31*(5), 763-770.
- [27] Fowler, J. S.; Volkow, N. D.; Logan, J.; Wang, G. J.; MacGregor, R. R.; Schlyer, D.; Wolf, A. P.; Pappas, N.; Alexoff, D.; Shea, C.; et al. Slow recovery of human brain MAO B after L-deprenyl (Selegiline) withdrawal. *Synapse*, **1994**, *18*(2), 86-93.
- [28] Arnett, C. D.; Fowler, J. S.; MacGregor, R. R.; Schlyer, D. J.; Wolf, A. P.; Langstrom, B.; Halldin, C. Turnover of brain monoamine oxidase measured *in vivo* by positron emission tomography using L-[¹¹C]deprenyl. *J. Neurochem.*, **1987**, *49*(2), 522-527.
- [29] Jahng, J. W.; Houpt, T. A.; Wessel, T. C.; Chen, K.; Shih, J. C.; Joh, T. H. Localization of monoamine oxidase A and B mRNA in the rat brain by *in situ* hybridization. *Synapse*, **1997**, *25*(1), 30-36.
- [30] Luque, J. M.; Kwan, S. W.; Abell, C. W.; Da Prada, M.; Richards, J. G. Cellular expression of mRNAs encoding monoamine oxidases A and B in the rat central nervous system. *J. Comp. Neurol.*, **1995**, *363*(4), 665-680.
- [31] Libert, S.; Pointer, K.; Bell, E. L.; Das, A.; Cohen, D. E.; Asara, J. M.; Kapur, K.; Bergmann, S.; Preisig, M.; Otowa, T.; Kendler, K. S.; Chen, X.; Hettema, J. M.; van den Oord, E. J.; Rubio, J. P.; Guarente, L. SIRT1 activates MAO-A in the brain to mediate anxiety and exploratory drive. *Cell*, **2011**, *147* (7), 1459-1472.
- [32] Saura, J.; Bleuel, Z.; Ulrich, J.; Mendelowitsch, A.; Chen, K.; Shih, J. C.; Malherbe, P.; Da Prada, M.; Richards, J. G. Molecular neuroanatomy of human monoamine oxidases A and B revealed by quantitative enzyme radioautography and *in situ* hybridization histochemistry. *Neuroscience*, **1996**, *70*(3), 755-774.
- [33] Westlund, K.N.; Denney, R.M.; Rose, R.M.; Abell, C.W. Localization of distinct monoamine oxidase A and monoamine oxidase B cell populations in human brainstem. *Neuroscience*, **1988**, *25*(2), 439-456.
- [34] Willoughby, J.; Glover, V.; Sandler, M. Histochemical localisation of monoamine oxidase A and B in rat brain. *J. Neural. Transm.*, **1988**, *74*(1), 29-42.
- [35] Shih, J.C.; Chen, K.; Ridd, M. J. Monoamine oxidase: from genes to behavior. *Annu. Rev. Neurosci.*, **1999**, *22*, 197-217.
- [36] Riederer, P.; Konrad, C.; Schay, V.; Kienzl, E.; Birkmayer, G.; Danielczyk, W.; Sofic, E.; Youdim, M. B. Localization of MAO-A and MAO-B in human brain: a step in understanding the therapeutic action of L-deprenyl. *Adv. Neurol.*, **1987**, *45*, 111-118.
- [37] Masson-Pevet, M.; Pevet, P. Cytochemical localization of type-A and -B monoamine oxidase in the rat pineal gland. *Cell Tissue Res.*, **1989**, *255*(2), 299-305.
- [38] O'Carroll, A. M.; Fowler, C. J.; Phillips, J. P.; Tobbia, I.; Tipton, K.F. The deamination of dopamine by human brain monoamine oxidase. Specificity for the two enzyme forms in seven brain regions. *Naunyn-Schmiedebergs Arch. Pharmacol.*, **1983**, *322*(3), 198-202.
- [39] Glover, V.; Sandler, M.; Owen, F.; Riley, G. J. Dopamine is a monoamine oxidase B substrate in man. *Nature*, **1977**, *265*(5589), 80-81.
- [40] Waldmeier, P. C.; Delini-Stula, A.; Maitre, L. Preferential deamination of dopamine by an A type monoamine oxidase in rat brain. *Naunyn-Schmiedebergs Arch. Pharmacol.*, **1976**, *292*(1), 9-14.
- [41] Egashira, T.; Yamamoto, T.; Yamanaka, Y. Some interrelated properties of A and B form monoamine oxidase in monkey brain mitochondria. *Jpn. J. Pharmacol.*, **1984**, *34*(3), 327-334.
- [42] Garrick, N. A.; Murphy, D. L. Species differences in the deamination of dopamine and other substrates for monoamine oxidase in brain. *Psychopharmacology (Berl)*, **1980**, *72*(1), 27-33.
- [43] Yu, P. H.; Hertz, L. Differential expression of type A and type B monoamine oxidase of mouse astrocytes in primary cultures. *J. Neurochem.*, **1982**, *39*(5), 1492-1495.
- [44] Youdim, M. B.; Edmondson, D.; Tipton, K. F. The therapeutic potential of monoamine oxidase inhibitors. *Nat. Rev. Neurosci.*, **2006**, *7*(4), 295-309.
- [45] Inyushin, M. Y.; Huertas, A.; Kucheryavykh, Y. V.; Kucheryavykh, L. Y.; Tsydzik, V.; Sanabria, P.; Eaton, M. J.; Skatchkov, S. N.; Rojas, L. V.; Wessinger, W. D. L-DOPA Uptake in Astrocytic Endfeet Enwrapping Blood Vessels in Rat Brain. *Parkinsons Dis.*, **2012**, *2012*, 321406.
- [46] Naoi, M.; Maruyama, W. Monoamine Oxidase Inhibitors as Neuroprotective Agents in Age-Dependent Neurodegenerative Disorders. *Curr. Pharm. Des.*, **2010**, *16*, 2799-2817.
- [47] Coppen, A. The biochemistry of affective disorders. *Br. J. Psychiatry*, **1967**, *113*(504), 1237-1264.
- [48] Schildkraut, J.J. The catecholamine hypothesis of affective disorders: a review of supporting evidence. *Am. J. Psychiatry*, **1965**, *122*(5), 509-522.
- [49] Bach, A.W.; Lan, N. C.; Johnson, D. L.; Abell, C. W.; Bembek, M. E.; Kwan, S. W.; Seeburg, P. H.; Shih, J. C. cDNA cloning of human liver monoamine oxidase A and B; molecular basis of differences in enzymatic properties. *Proc. Natl. Acad. Sci. U S A*, **1988**, *85*(13), 4934-4938.
- [50] Derry, J. M.; Lan, N. C.; Shih, J. C.; Barnard, E. A.; Barnard, P. J. Localization of monoamine oxidase A and B genes on the mouse X chromosome. *Nucleic Acids Res.*, **1989**, *17*(20), 8403.
- [51] Lan, N.C.; Heinzmann, C.; Gal, A.; Klisak, I.; Orth, U.; Lai, E.; Grimsby, J.; Sparkes, R. S.; Mohandas, T.; Shih, J. C. Human monoamine oxidase A and B genes map to Xp 11.23 and are deleted in a patient with Norrie disease. *Genomics*, **1989**, *4*(4), 552-559.
- [52] Grimsby, J.; Chen, K.; Wang, L. J.; Lan, N. C.; Shih, J. C. Human monoamine oxidase A and B genes exhibit identical exon-intron organization. *Proc. Natl. Acad. Sci. U S A*, **1991**, *88*(9), 3637-3641.
- [53] Costa-Mallen, P.; Kelada, S. N.; Costa, L. G.; Checkoway, H. Characterization of the *in vitro* transcriptional activity of polymorphic alleles of the human monoamine oxidase-B gene. *Neurosci. Lett.*, **2005**, *383*(1-2), 171-175.
- [54] Sabol, S. Z.; Hu, S.; Hamer, D. A functional polymorphism in the monoamine oxidase A gene promoter. *Hum. Genet.*, **1998**, *103*(3), 273-279.
- [55] Hotamisligil, G.S.; Breakefield, X.O. Human monoamine oxidase A gene determines levels of enzyme activity. *Am. J. Hum. Genet.*, **1991**, *49*(2), 383-392.
- [56] Hotamisligil, G.S.; Girman, A.S.; Fink, J.S.; Tivol, E.; Shalish, C.; Trofatter, J.; Baenziger, J.; Diamond, S.; Markham, C.; Sullivan, J.; et al. Hereditary variations in monoamine oxidase as a risk factor for Parkinson's disease. *Mov. Disord.*, **1994**, *9*(3), 305-310.
- [57] Wu, Y.H.; Fischer, D.F.; Swaab, D.F. A promoter polymorphism in the monoamine oxidase A gene is associated with the pineal MAOA activity in Alzheimer's disease patients. *Brain Res.*, **2007**, *1167*, 13-19.
- [58] Huang, Y. Y.; Cate, S. P.; Battistuzzi, C.; Oquendo, M. A.; Brent, D.; Mann, J. J. An association between a functional polymorphism

- in the monoamine oxidase a gene promoter, impulsive traits and early abuse experiences. *Neuropsychopharmacology*, **2004**, *29*(8), 1498-1505.
- [59] Gade, R.; Muhleman, D.; Blake, H.; MacMurray, J.; Johnson, P.; Verde, R.; Saucier, G.; Comings, D. E. Correlation of length of VNTR alleles at the X-linked MAOA gene and phenotypic effect in Tourette syndrome and drug abuse. *Mol. Psychiatry*, **1998**, *3*(1), 50-60.
- [60] Kunugi, H.; Ishida, S.; Kato, T.; Tatsumi, M.; Sakai, T.; Hattori, M.; Hirose, T.; Nanko, S. A functional polymorphism in the promoter region of monoamine oxidase-A gene and mood disorders. *Mol. Psychiatry*, **1999**, *4*(4), 393-395.
- [61] Meyer, J. H.; Ginovart, N.; Boovariwala, A.; Sagrati, S.; Hussey, D.; Garcia, A.; Young, T.; Prashak-Rieder, N.; Wilson, A. A.; Houle, S. Elevated monoamine oxidase a levels in the brain: an explanation for the monoamine imbalance of major depression. *Arch. Gen. Psychiatry*, **2006**, *63*(11), 1209-1216.
- [62] Sacher, J.; Wilson, A.A.; Houle, S.; Rusjan, P.; Hassan, S.; Bloomfield, P. M.; Stewart, D. E.; Meyer, J. H. Elevated brain monoamine oxidase A binding in the early postpartum period. *Arch. Gen. Psychiatry*, **2010**, *67*(5), 468-474.
- [63] Bacher, I.; Houle, S.; Xu, X.; Zawertailo, L.; Soliman, A.; Wilson, A. A.; Selby, P.; George, T. P.; Sacher, J.; Miler, L.; Kish, S. J.; Rusjan, P.; Meyer, J. H. Monoamine oxidase A binding in the prefrontal and anterior cingulate cortices during acute withdrawal from heavy cigarette smoking. *Arch. Gen. Psychiatry*, **2011**, *68*(8), 817-826.
- [64] Essman, W.B. Serotonin and monoamine oxidase in mouse skin: effects of cigarette smoke exposure. *J. Med.*, **1977**, *8*(2), 95-101.
- [65] Herrmann, M.; King, K.; Weitzman, M. Prenatal tobacco smoke and postnatal secondhand smoke exposure and child neuro development. *Curr. Opin. Pediatr.*, **2008**, *20*(2), 184-190.
- [66] Weissman, M. M.; Warner, V.; Wickramaratne, P. J.; Kandel, D. B. Maternal smoking during pregnancy and psychopathology in offspring followed to adulthood. *J. Am. Acad. Child Adolesc. Psychiatry*, **1999**, *38*(7), 892-899.
- [67] Rasanen, P.; Hakko, H.; Isohanni, M.; Hodgins, S.; Jarvelin, M. R.; Tiihonen, J. Maternal smoking during pregnancy and risk of criminal behavior among adult male offspring in the Northern Finland 1966 Birth Cohort. *Am. J. Psychiatry*, **1999**, *156*(6), 857-862.
- [68] Brennan, P. A.; Grekin, E. R.; Mortensen, E. L.; Mednick, S. A. Relationship of maternal smoking during pregnancy with criminal arrest and hospitalization for substance abuse in male and female adult offspring. *Am. J. Psychiatry*, **2002**, *159*(1), 48-54.
- [69] Fowler, J. S.; Volkow, N. D.; Wang, G. J.; Pappas, N.; Logan, J.; MacGregor, R.; Alexoff, D.; Wolf, A. P.; Warner, D.; Cilento, R.; Zezulkova, I. Neuropharmacological actions of cigarette smoke: brain monoamine oxidase B (MAO B) inhibition. *J. Addict. Dis.*, **1998**, *17*(1), 23-34.
- [70] Costa-Mallen, P.; Costa, L. G.; Checkoway, H. Genotype combinations for monoamine oxidase-B intron 13 polymorphism and dopamine D2 receptor Taq1B polymorphism are associated with ever-smoking status among men. *Neurosci. Lett.*, **2005**, *385*(2), 158-162.
- [71] Leroy, C.; Bragulat, V.; Berlin, I.; Gregoire, M. C.; Bottlaender, M.; Roumenov, D.; Dolle, F.; Bourgeois, S.; Penttila, J.; Artiges, E.; Martinot, J. L.; Trichard, C. Cerebral monoamine oxidase A inhibition in tobacco smokers confirmed with PET and [11C]befloxatone. *J. Clin. Psychopharmacol.*, **2009**, *29*(1), 86-88.
- [72] Birkmayer, W.; Riederer, P.; Youdim, M. B.; Linauer, W. The potentiation of the anti aknetic effect after L-dopa treatment by an inhibitor of MAO-B, Deprenil. *J. Neural Transm.*, **1975**, *36*(3-4), 303-326.
- [73] Launay, J.M.; Del Pino, M.; Chironi, G.; Callebert, J.; Peoc'h, K.; Megniel, J.L.; Mallet, J.; Simon, A.; Rendu, F. Smoking induces long-lasting effects through a monoamine-oxidase epigenetic regulation. *PLoS One*, **2009**, *4*(11), e7959.
- [74] Shumay, E.; Fowler, J.S. Identification and characterization of putative methylation targets in the MAOA locus using bioinformatic approaches. *Epigenetics*, **2010**, *5*(4), 325-342.
- [75] Pinsonneault, J.K.; Papp, A.C.; Sadee, W. Allelic mRNA expression of X-linked monoamine oxidase a (MAOA) in human brain: dissection of epigenetic and genetic factors. *Hum. Mol. Genet.*, **2006**, *15*(17), 2636-2649.
- [76] Philibert, R. A.; Gunter, T. D.; Beach, S. R.; Brody, G. H.; Madan, A. MAOA methylation is associated with nicotine and alcohol dependence in women. *Am. J. Med. Genet. B Neuropsychiatr. Genet.*, **2008**, *147B*(5), 565-570.
- [77] Mimasu, S.; Sengoku, T.; Fukuzawa, S.; Umehara, T.; Yokoyama, S. Crystal structure of histone demethylase LSD1 and tranlycyp romine at 2.25 Å. *Biochem. Biophys. Res. Commun.*, **2008**, *366*(1), 15-22.
- [78] Hampp, G.; Ripperger, J. A.; Houben, T.; Schmutz, I.; Blex, C.; Perreau-Lenz, S.; Brunk, I.; Spanagel, R.; Ahnert-Hilger, G.; Meijer, J. H.; Albrecht, U. Regulation of monoamine oxidase A by circadian-clock components implies clock influence on mood. *Curr. Biol.*, **2008**, *18*(9), 678-683.
- [79] Raddatz, R.; Parini, A.; Lanier, S. M. Imidazoline/guanidinium binding domains on monoamine oxidases. Relationship to subtypes of imidazoline-binding proteins and tissue-specific interaction of imidazoline ligands with monoamine oxidase B. *J. Biol. Chem.*, **1995**, *270*(46), 27961-27968.
- [80] Cases, O.; Seif, I.; Grimsby, J.; Gaspar, P.; Chen, K.; Pourmin, S.; Muller, U.; Aguet, M.; Babinet, C.; Shih, J. C.; et al. Aggressive behavior and altered amounts of brain serotonin and norepinephrine in mice lacking MAOA. *Science*, **1995**, *268*(5218), 1763-1766.
- [81] Kearney, E. B.; Salach, J. I.; Walker, W. H.; Seng, R. L.; Kenney, W.; Zeszotek, E.; Singer, T. P. The covalently-bound flavin of hepatic monoamine oxidase. 1. Isolation and sequence of a flavin peptide and evidence for binding at the 8α position. *Eur. J. Biochem.*, **1971**, *24*(2), 321-327.
- [82] Hiro, I.; Tsugeno, Y.; Hirashiki, I.; Ogata, F.; Ito, A. Characterization of rat monoamine oxidase A with noncovalently-bound FAD expressed in yeast cells. *J. Biochem.*, **1996**, *120*(4), 759-765.
- [83] Nandigama, R. K.; Edmondson, D. E. Influence of FAD structure on its binding and activity with the C406A mutant of recombinant human liver monoamine oxidase A. *J. Biol. Chem.*, **2000**, *275*(27), 20527-20532.
- [84] Geha, R. M.; Rebrin, I.; Chen, K.; Shih, J. C. Substrate and inhibitor specificities for human monoamine oxidase A and B are influenced by a single amino acid. *J. Biol. Chem.*, **2001**, *276*(13), 9877-9882.
- [85] Wei, Z.; Satram-Maharaj, T.; Chaharyn, B.; Kuski, K.; Pennington, P. R.; Cao, X.; Chlan, J.; Mousseau, D. D. Aspartic acid substitutions in monoamine oxidase-A reveal both catalytic-dependent and -independent influences on cell viability and proliferation. *J. Neural Transm.*, **2012**, *119*(11), 1285-94.
- [86] Chiou, S. H.; Ku, H. H.; Tsai, T. H.; Lin, H. L.; Chen, L. H.; Chien, C. S.; Ho, L. L.; Lee, C. H.; Chang, Y. L. Moclobemide upregulated Bcl-2 expression and induced neural stem cell differentiation into serotonergic neuron via extracellular-regulated kinase pathway. *Br. J. Pharmacol.*, **2006**, *148*(5), 587-598.
- [87] Inaba-Hasegawa, K.; Akao, Y.; Maruyama, W.; Naoi, M. Type A monoamine oxidase is associated with induction of neuroprotective Bcl-2 by rasagiline, an inhibitor of type B monoamine oxidase. *J. Neural Transm.*, **2012**, *119*(4), 405-414.
- [88] Saura, J.; Luque, J. M.; Cesura, A. M.; Da Prada, M.; Chan-Palay, V.; Huber, G.; Löffler, J.; Richards, J. G. Increased monoamine oxidase B activity in plaque-associated astrocytes of Alzheimer brains revealed by quantitative enzyme radioautography. *Neuroscience*, **1994**, *62*(1), 15-30.
- [89] Grunblatt, E.; Schlosser, R.; Fischer, P.; Fischer, M. O.; Li, J.; Koutsilieris, E.; Wichart, I.; Sterba, N.; Rujescu, D.; Moller, H. J.; Adamcyk, W.; Dittrich, B.; Muller, F.; Oberegger, K.; Gatterer, G.; Jellinger, K. J.; Mostafaie, N.; Jungwirth, S.; Huber, K.; Tragl, K. H.; Danielczyk, W.; Riederer, P. Oxidative stress related markers in the "VITA" and the centenarian projects. *Neurobiol. Aging*, **2005**, *26*(4), 429-438.
- [90] Adolfsson, R.; Gottfries, C. G.; Oreland, L.; Wiberg, A.; Winblad, B. Increased activity of brain and platelet monoamine oxidase in dementia of Alzheimer type. *Life Sci.*, **1980**, *27*(12), 1029-1034.
- [91] Parnetti, L.; Reboldi, G. P.; Santucci, C.; Santucci, A.; Gaiti, A.; Brunetti, M.; Cecchetti, R.; Senin, U. Platelet MAO-B activity as a marker of behavioural characteristics in dementia disorders. *Aging (Milano)*, **1994**, *6*(3), 201-207.
- [92] Reinikainen, K. J.; Paljarvi, L.; Halonen, T.; Malminen, O.; Kosma, V. M.; Laakso, M.; Riekkinen, P. J. Dopaminergic system and monoamine oxidase-B activity in Alzheimer's disease. *Neurobiol. Aging*, **1988**, *9*(3), 245-252.

- [93] Emilsson, L.; Saetre, P.; Balciuniene, J.; Castensson, A.; Cairns, N.; Jazin, E. E. Increased monoamine oxidase messenger RNA expression levels in frontal cortex of Alzheimer's disease patients. *Neurosci Lett.*, **2002**, *326* (1), 56-60.
- [94] Nakamura, S.; Kawamata, T.; Akiguchi, I.; Kameyama, M.; Nakamura, N.; Kimura, H. Expression of monoamine oxidase B activity in astrocytes of senile plaques. *Acta Neuropathol. (Berl)*, **1990**, *80* (4), 419-425.
- [95] Kennedy, B. P.; Ziegler, M. G.; Alford, M.; Hansen, L. A.; Thal, L. J.; Masliah, E. Early and persistent alterations in prefrontal cortex MAO A and B in Alzheimer's disease. *J. Neural. Transm.*, **2003**, *110* (7), 789-801.
- [96] Sparks, D. L.; Woeltz, V. M.; Markesbery, W. R. Alterations in brain monoamine oxidase activity in aging, Alzheimer's disease, and Pick's disease. *Arch. Neurol.*, **1991**, *48* (7), 718-721.
- [97] Sherif, F.; Gottfries, C. G.; Alafuzoff, I.; Oreland, L. Brain γ -aminobutyrate aminotransferase (GABA-T) and monoamine oxidase (MAO) in patients with Alzheimer's disease. *J. Neural Transm. Park. Dis. Dement. Sect.*, **1992**, *4*(3), 227-240.
- [98] Chan-Palay, V.; Hochli, M.; Savaskan, E.; Hungerecker, G. Calbindin D-28k and monoamine oxidase A immunoreactive neurons in the nucleus basalis of Meynert in senile dementia of the Alzheimer type and Parkinson's disease. *Dementia*, **1993**, *4*(1), 1-15.
- [99] Green, A. R.; Mitchell, B. D.; Tordoff, A. F.; Youdim, M. B. Evidence for dopamine deamination by both type A and type B monoamine oxidase in rat brain *in vivo* and for the degree of inhibition of enzyme necessary for increased functional activity of dopamine and 5-hydroxytryptamine. *Br. J. Pharmacol.*, **1977**, *60*(3), 343-349.
- [100] Sacher, J.; Houle, S.; Parkes, J.; Rusjan, P.; Sagrati, S.; Wilson, A. A.; Meyer, J. H. Monoamine oxidase A inhibitor occupancy during treatment of major depressive episodes with moclobemide or St. John's wort: an [¹¹C]-harmine PET study. *J. Psychiatry Neurosci.*, **2011**, *36*(6), 375-382.
- [101] Burke, W. J.; Li, S. W.; Schmitt, C. A.; Xia, P.; Chung, H. D.; Gillespie, K. N. Accumulation of 3,4-dihydroxyphenylglycolaldehyde, the neurotoxic monoamine oxidase A metabolite of norepinephrine, in locus ceruleus cell bodies in Alzheimer's disease: mechanism of neuron death. *Brain Res.*, **1999**, *816*(2), 633-637.
- [102] Burke, W. J.; Li, S. W.; Chung, H. D.; Ruggiero, D. A.; Kristal, B. S.; Johnson, E. M.; Lampe, P.; Kumar, V. B.; Franko, M.; Williams, E. A.; Zahm, D. S. Neurotoxicity of MAO metabolites of catecholamine neurotransmitters: role in neurodegenerative diseases. *Neurotoxicology*, **2004**, *25*(1-2), 101-115.
- [103] Grudzien, A.; Shaw, P.; Weintraub, S.; Bigio, E.; Mash, D. C.; Mesulam, M. M. Locus coeruleus neurofibrillary degeneration in aging, mild cognitive impairment and early Alzheimer's disease. *Neurobiol. Aging*, **2007**, *28* (3), 327-335.
- [104] Marcyniuk, B.; Mann, D. M.; Yates, P. O. Loss of nerve cells from locus coeruleus in Alzheimer's disease is topographically arranged. *Neurosci. Lett.*, **1986**, *64*(3), 247-252.
- [105] Parvizi, J.; Van Hoesen, G.W.; Damasio, A. The selective vulnerability of brainstem nuclei to Alzheimer's disease. *Ann. Neurol.*, **2001**, *49*(1), 53-66.
- [106] Rub, U.; Del Tredici, K.; Schultz, C.; Thal, D. R.; Braak, E.; Braak, H. The evolution of Alzheimer's disease-related cytoskeletal pathology in the human raphe nuclei. *Neuropathol. Appl. Neurobiol.*, **2000**, *26*(6), 553-567.
- [107] Zweig, R.M.; Ross, C. A.; Hedreen, J. C.; Steele, C.; Cardillo, J. E.; Whitehouse, P. J.; Folstein, M. F.; Price, D. L. The neuropathology of aminergic nuclei in Alzheimer's disease. *Ann. Neurol.*, **1988**, *24*(2), 233-242.
- [108] Gundlach, C.; Lu, N. Z.; Bethea, C. L. Ovarian steroid regulation of monoamine oxidase-A and -B mRNAs in the macaque dorsal raphe and hypothalamic nuclei. *Psychopharmacology (Berl)*, **2002**, *160* (3), 271-282.
- [109] Holschneider, D.P.; Kumazawa, T.; Chen, K.; Shih, J. C. Tissue-specific effects of estrogen on monoamine oxidase A and B in the rat. *Life Sci.*, **1998**, *63*(3), 155-160.
- [110] Ou, X. M.; Chen, K.; Shih, J. C. Monoamine oxidase A and repressor R1 are involved in apoptotic signaling pathway. *Proc. Natl. Acad. Sci. U S A*, **2006**, *103* (29), 10923-10928.
- [111] Fitzgerald, J. C.; Ufer, C.; Billett, E. E. A link between monoamine oxidase-A and apoptosis in serum deprived human SH-SY5Y neuroblastoma cells. *J. Neural Transm.*, **2007**, *114*(6), 807-810.
- [112] Wang, C. C.; Borchert, A.; Ugun-Klusek, A.; Tang, L. Y.; Lui, W. T.; Chu, C. Y.; Billett, E.; Kuhn, H.; Ufer, C. Monoamine oxidase A expression is vital for embryonic brain development by modulating developmental apoptosis. *J. Biol. Chem.*, **2011**, *286*(32), 28322-28330.
- [113] Bianchi, P.; Seguelas, M. H.; Parini, A.; Cambon, C. Activation of pro-apoptotic cascade by dopamine in renal epithelial cells is fully dependent on hydrogen peroxide generation by monoamine oxidases. *J. Am. Soc. Nephrol.*, **2003**, *14*(4), 855-862.
- [114] Yi, H.; Akao, Y.; Maruyama, W.; Chen, K.; Shih, J.; Naoi, M. Type A monoamine oxidase is the target of an endogenous dopaminergic neurotoxin, N-methyl(R)salsolinol, leading to apoptosis in SH-SY5Y cells. *J. Neurochem.*, **2006**, *96*(2), 541-549.
- [115] Saura, J.; Richards, J. G.; Mahy, N. Differential age-related changes of MAO-A and MAO-B in mouse brain and peripheral organs. *Neurobiol. Aging*, **1994**, *15*(4), 399-408.
- [116] Saura, J.; Richards, J. G.; Mahy, N. Age-related changes on MAO in BL/C57 mouse tissues: a quantitative radioautographic study. *J. Neural Transm. Suppl.*, **1994**, *41*, 89-94.
- [117] Mousseau, D. D.; McManus, D. J.; Baker, G. B.; Juorio, A. V.; Dewhurst, W. G.; Greenshaw, A. J. Effects of age and of chronic antidepressant treatment on [3H]tryptamine and [3H]dihydroalprenolol binding to rat cortical membranes. *Cell. Mol. Neurobiol.*, **1993**, *13*(1), 3-13.
- [118] Rybaczyk, L. A.; Bashaw, M. J.; Pathak, D. R.; Huang, K. An indicator of cancer: downregulation of monoamine oxidase-A in multiple organs and species. *BMC Genomics*, **2008**, *9*, 134.
- [119] Lizcano, J. M.; Escrich, E.; Ribalta, T.; Muntane, J.; Unzeta, M. Amine oxidase activities in rat breast cancer induced experimentally with 7,12-dimethylbenz(α)anthracene. *Biochem. Pharmacol.*, **1991**, *42*(2), 263-269.
- [120] Pai, V. P.; Marshall, A. M.; Hernandez, L. L.; Buckley, A. R.; Horseman, N. D. Altered serotonin physiology in human breast cancers favors paradoxical growth and cell survival. *Breast Cancer Res.*, **2009**, *11*(6), R81.
- [121] Peehl, D. M.; Coram, M.; Khine, H.; Reese, S.; Nolley, R.; Zhao, H. The significance of monoamine oxidase-A expression in high grade prostate cancer. *J. Urol.*, **2008**, *180*(5), 2206-2211.
- [122] Yu, D. S.; Hsieh, D. S.; Chen, H. I.; Chang, S. Y. The expression of neuropeptides in hyperplastic and malignant prostate tissue and its possible clinical implications. *J. Urol.*, **2001**, *166*(3), 871-875.
- [123] Jungwirth, N.; Haeblerle, L.; Schrott, K. M.; Wullich, B.; Krause, F. S. Serotonin used as prognostic marker of urological tumors. *World J. Urol.*, **2008**, *26*(5), 499-504.
- [124] Wei, Z.; Gabriel, G.G.; Rui, L.; Cao, X.; Pennington, P. R.; Chlan-Fourney, J.; Nazarali, A. J.; Baker, G. B.; Mousseau, D. D. Monoamine oxidase-A physically interacts with presenilin-1 (M146V) in the mouse cortex. *J. Alzheimers Dis.*, **2012**, *28*(2), 403-422.
- [125] Louvi, A.; Sisodia, S. S.; Grove, E. A. Presenilin 1 in migration and morphogenesis in the central nervous system. *Development*, **2004**, *131*(13), 3093-3105.
- [126] Wines-Samuels, M.; Handler, M.; Shen, J. Role of presenilin-1 in cortical lamination and survival of Cajal-Retzius neurons. *Dev. Biol.*, **2005**, *277*(2), 332-346.
- [127] Cases, O.; Vitalis, T.; Seif, I.; De Maeyer, E.; Sotelo, C.; Gaspar, P. Lack of barrels in the somatosensory cortex of monoamine oxidase A-deficient mice: role of a serotonin excess during the critical period. *Neuron*, **1996**, *16*(2), 297-307.
- [128] Scott, A. L.; Bortolato, M.; Chen, K.; Shih, J. C. Novel monoamine oxidase A knock out mice with human-like spontaneous mutation. *Neuroreport*, **2008**, *19* (7), 739-743.
- [129] Cheng, A.; Scott, A.L.; Ladenheim, B.; Chen, K.; Ouyang, X.; Lathia, J. D.; Mughal, M.; Cadet, J. L.; Mattson, M. P.; Shih, J. C. Monoamine oxidases regulate telencephalic neural progenitors in late embryonic and early postnatal development. *J. Neurosci.*, **2010**, *30*(32), 10752-10762.
- [130] Brunner, H. G.; Nelen, M.; Breakefield, X. O.; Ropers, H. H.; van Oost, B. A. Abnormal behavior associated with a point mutation in the structural gene for monoamine oxidase A. *Science*, **1993**, *262* (5133), 578-580.

- [131] Chau, K. Y.; Cooper, J. M.; Schapira, A. H. Rasagiline protects against α -synuclein induced sensitivity to oxidative stress in dopaminergic cells. *Neurochem. Int.*, **2010**, *57* (5), 525-529.
- [132] Rodriguez, S.; Ito, T.; He, X. J.; Uchida, K.; Nakayama, H. Resistance of the golden hamster to 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-neurotoxicity is not only related with low levels of cerebral monoamine oxidase-B. *Exp. Toxicol. Pathol.*, **2011**, *65*(1-2), 127-133.
- [133] Mandel, S. A.; Sagi, Y.; Amit, T. Rasagiline promotes regeneration of substantia nigra dopaminergic neurons in post-MPTP-induced Parkinsonism via activation of tyrosine kinase receptor signaling pathway. *Neurochem. Res.*, **2007**, *32*(10), 1694-1699.
- [134] Tatton, W. G.; Chalmers-Redman, R. M. Modulation of gene expression rather than monoamine oxidase inhibition: (-)-deprenyl-related compounds in controlling neurodegeneration. *Neurology*, **1996**, *47* (6 Suppl 3), S171-183.
- [135] Tatton, W. G.; Wadia, J. S.; Ju, W. Y.; Chalmers-Redman, R. M.; Tatton, N. A. (-)-Deprenyl reduces neuronal apoptosis and facilitates neuronal outgrowth by altering protein synthesis without inhibiting monoamine oxidase. *J. Neural Transm. Suppl.*, **1996**, *48*, 45-59.
- [136] Tsugeno, Y.; Hirashiki, I.; Ogata, F.; Ito, A. Regions of the molecule responsible for substrate specificity of monoamine oxidase A and B: a chimeric enzyme analysis. *J. Biochem. (Tokyo)*, **1995**, *118*(5), 974-980.
- [137] De Colibus, L.; Li, M.; Binda, C.; Lustig, A.; Edmondson, D. E.; Mattevi, A. Three-dimensional structure of human monoamine oxidase A (MAO A): relation to the structures of rat MAO A and human MAO B. *Proc. Natl. Acad. Sci. U S A*, **2005**, *102*(36), 12684-12689.
- [138] Rebrin, I.; Geha, R. M.; Chen, K.; Shih, J. C. Effects of carboxyl-terminal truncations on the activity and solubility of human monoamine oxidase B. *J. Biol. Chem.*, **2001**, *276*(31), 29499-29506.
- [139] Weyler, W. Functional expression of C-terminally truncated human monoamine oxidase type A in *Saccharomyces cerevisiae*. *J. Neural Transm. Suppl.*, **1994**, *41*, 3-15.
- [140] Chen, K.; Wu, H. F.; Shih, J. C. Influence of C terminus on monoamine oxidase A and B catalytic activity. *J. Neurochem.*, **1996**, *66*(2), 797-803.
- [141] Maycock, A.L.; Abeles, R. H.; Salach, J. I.; Singer, T. P. The structure of the covalent adduct formed by the interaction of 3-dimethylamino-1-propyne and the flavine of mitochondrial amine oxidase. *Biochemistry*, **1976**, *15*(1), 114-125.
- [142] Gareri, P.; Falconi, U.; De Fazio, P.; De Sarro, G. Conventional and new antidepressant drugs in the elderly. *Prog. Neurobiol.*, **2000**, *61*(4), 353-396.
- [143] Rosenzweig, P.; Patat, A.; Zieleniuk, I.; Cimarosti, I.; Allain, H.; Gandon, J. M. Cognitive performance in elderly subjects after a single dose of bexlofatone, a new reversible selective monoamine oxidase A inhibitor. *Clin. Pharmacol. Ther.*, **1998**, *64*(2), 211-222.
- [144] Fowler, J. S.; Logan, J.; Azzaro, A. J.; Fielding, R. M.; Zhu, W.; Poshusta, A. K.; Burch, D.; Brand, B.; Free, J.; Asgharnejad, M.; Wang, G. J.; Telang, F.; Hubbard, B.; Jayne, M.; King, P.; Carter, P.; Carter, S.; Xu, Y.; Shea, C.; Muench, L.; Alexoff, D.; Shumay, E.; Schueller, M.; Warner, D.; Apelskog-Torres, K. Reversible inhibitors of monoamine oxidase-A (RIMAs): robust, reversible inhibition of human brain MAO-A by CX157. *Neuropsychopharmacology*, **2010**, *35*(3), 623-631.
- [145] Nishimura, A. L.; Guindalini, C.; Oliveira, J. R.; Nitrini, R.; Bahia, V. S.; de Brito-Marques, P. R.; Otto, P. A.; Zatz, M. Monoamine oxidase a polymorphism in Brazilian patients: risk factor for late-onset Alzheimer's disease? *J. Mol. Neurosci.*, **2005**, *27*(2), 213-217.
- [146] Takehashi, M.; Tanaka, S.; Masliah, E.; Ueda, K. Association of monoamine oxidase A gene polymorphism with Alzheimer's disease and Lewy body variant. *Neurosci. Lett.*, **2002**, *327*(2), 79-82.
- [147] Caraci, F.; Copani, A.; Nicoletti, F.; Drago, F. Depression and Alzheimer's disease: neurobiological links and common pharmacological targets. *Eur. J. Pharmacol.*, **2010**, *626*(1), 64-71.
- [148] Geerlings, M. I.; den Heijer, T.; Koudstaal, P. J.; Hofman, A.; Breteler, M. M. History of depression, depressive symptoms, and medial temporal lobe atrophy and the risk of Alzheimer disease. *Neurology*, **2008**, *70*(15), 1258-1264.
- [149] Wuwongse, S.; Chang, R. C.; Law, A. C. The putative neurodegenerative links between depression and Alzheimer's disease. *Prog. Neurobiol.*, **2010**, *91*(4), 362-375.
- [150] Malorni, W.; Giammarioli, A. M.; Matarrese, P.; Pietrangeli, P.; Agostinelli, E.; Ciaccio, A.; Grassilli, E.; Mondovi, B. Protection against apoptosis by monoamine oxidase A inhibitors. *FEBS Lett.*, **1998**, *426* (1), 155-159.
- [151] Maher, P.; Davis, J. B. The role of monoamine metabolism in oxidative glutamate toxicity. *J. Neurosci.*, **1996**, *16* (20), 6394-6401.
- [152] Maragos, W. F.; Young, K. L.; Altman, C. S.; Pocernich, C. B.; Drake, J.; Butterfield, D. A.; Seif, I.; Holschneider, D. P.; Chen, K.; Shih, J. C. Striatal damage and oxidative stress induced by the mitochondrial toxin malonate are reduced in clorgyline-treated rats and MAO-A deficient mice. *Neurochem. Res.*, **2004**, *29* (4), 741-746.
- [153] Birks, J.; Flicker, L. Selegiline for Alzheimer's disease. *Cochrane Database Syst. Rev.*, **2003**, (1), CD000442.
- [154] Ebadi, M.; Brown-Borg, H.; Ren, J.; Sharma, S.; Shavali, S.; El ReFaey, H.; Carlson, E. C. Therapeutic efficacy of selegiline in neurodegenerative disorders and neurological diseases. *Curr. Drug Targets*, **2006**, *7* (11), 1513-1529.
- [155] Sterling, J.; Herzig, Y.; Goren, T.; Finkelstein, N.; Lerner, D.; Goldenberg, W.; Miskolczi, I.; Molnar, S.; Rantal, F.; Tamas, T.; Toth, G.; Zagyya, A.; Zekany, A.; Finberg, J.; Lavian, G.; Gross, A.; Friedman, R.; Razin, M.; Huang, W.; Kraiss, B.; Chorev, M.; Youdim, M. B.; Weinstock, M. Novel dual inhibitors of AChE and MAO derived from hydroxy aminoindan and phenethylamine as potential treatment for Alzheimer's disease. *J. Med. Chem.*, **2002**, *45*(24), 5260-5279.
- [156] Yogeve-Falach, M.; Bar-Am, O.; Amit, T.; Weinreb, O.; Youdim, M. B. A multifunctional, neuroprotective drug, ladostigil (TV3326), regulates holo-APP translation and processing. *Faseb J.*, **2006**, *20*(12), 2177-2179.
- [157] Veazey, C.; Aki, S. O.; Cook, K. F.; Lai, E. C.; Kunik, M. E. Prevalence and treatment of depression in Parkinson's disease. *J. Neuropsychiatry Clin. Neurosci.*, **2005**, *17*(3), 310-323.
- [158] Yamamoto, M. Depression in Parkinson's disease: its prevalence, diagnosis, and neurochemical background. *J. Neurol.*, **2001**, *248* Suppl 3, III5-11.
- [159] Hunter, K. R.; Boakes, A. J.; Laurence, D. R.; Stern, G. M. Monoamine oxidase inhibitors and L-dopa. *Br. Med. J.*, **1970**, *3*(5719), 388.
- [160] Sjoqvist, F. Psychotropic drugs (2). Interaction between monoamine oxidase (MAO) inhibitors and other substances. *Proc. R. Soc. Med.*, **1965**, *58*(11 Part 2), 967-978.
- [161] Colzi, A.; D'Agostini, F.; Cesura, A. M.; Borroni, E.; Da Prada, M. Monoamine oxidase-A inhibitors and dopamine metabolism in rat caudatus: evidence that an increased cytosolic level of dopamine displaces reversible monoamine oxidase-A inhibitors *in vivo*. *J. Pharmacol. Exp. Ther.*, **1993**, *265*(1), 103-111.
- [162] Davidson, J. R. Pharmacotherapy of social phobia. *Acta Psychiatr. Scand. Suppl.*, **2003**, (417), 65-71.
- [163] Bottlaender, M.; Dolle, F.; Guenther, I.; Roumenov, D.; Fuseau, C.; Bramouille, Y.; Curet, O.; Jegham, J.; Pinquier, J. L.; George, P.; Valette, H. Mapping the cerebral monoamine oxidase type A: positron emission tomography characterization of the reversible selective inhibitor [11C]bexlofatone. *J. Pharmacol. Exp. Ther.*, **2003**, *305*(2), 467-473.
- [164] Youdim, M. B.; Bakhle, Y. S. Monoamine oxidase: isoforms and inhibitors in Parkinson's disease and depressive illness. *Br. J. Pharmacol.*, **2006**, *147* Suppl 1, S287-296.
- [165] Haefely, W.; Burkard, W. P.; Cesura, A. M.; Kettler, R.; Lorez, H. P.; Martin, J. R.; Richards, J. G.; Scherschlicht, R.; Da Prada, M. Biochemistry and pharmacology of moclobemide, a prototype RIMA. *Psychopharmacology (Berl)*, **1992**, *106* Suppl, S6-14.
- [166] Youdim, M. B.; Weinstock, M. Therapeutic applications of selective and non-selective inhibitors of monoamine oxidase A and B that do not cause significant tyramine potentiation. *Neurotoxicology*, **2004**, *25*(1-2), 243-250.
- [167] Sternic, N.; Kacar, A.; Filipovic, S.; Svetel, M.; Kostic, V. S. The therapeutic effect of moclobemide, a reversible selective monoamine oxidase A inhibitor, in Parkinson's disease. *Clin. Neuropharmacol.*, **1998**, *21*(2), 93-96.
- [168] Holt, A.; Berry, M. D.; Boulton, A. A. On the binding of monoamine oxidase inhibitors to some sites distinct from the MAO

- active site, and effects thereby elicited. *Neurotoxicology*, **2004**, 25(1-2), 251-266.
- [169] Gillman, P. K. Advances pertaining to the pharmacology and interactions of irreversible nonselective monoamine oxidase inhibitors. *J. Clin. Psychopharmacol.*, **2011**, 31(1), 66-74.
- [170] Holt, A.; Baker, G. B. Inhibition of rat brain monoamine oxidase enzymes by fluoxetine and norfluoxetine. *Naunyn-Schmiedeberg's Arch. Pharmacol.*, **1996**, 354(1), 17-24.
- [171] Ono, K.; Hasegawa, K.; Naiki, H.; Yamada, M. Anti-Parkinsonian agents have anti-amyloidogenic activity for Alzheimer's β -amyloid fibrils *in vitro*. *Neurochem. Int.*, **2006**, 48(4), 275-285.
- [172] de Lima, M. N.; Laranja, D. C.; Caldana, F.; Bromberg, E.; Roessler, R.; Schroder, N. Reversal of age-related deficits in object recognition memory in rats with l-deprenyl. *Exp. Gerontol.*, **2005**, 40(6), 506-511.
- [173] Fuller, R. W.; Hemrick-Luecke, S. K. Mechanisms of MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) neurotoxicity to striatal dopamine neurons in mice. *Prog. Neuropsychopharmacol. Biol. Psychiatry*, **1985**, 9(5-6), 687-690.
- [174] Langston, J. W.; Irwin, I.; Langston, E. B.; Forno, L. S. Pargyline prevents MPTP-induced parkinsonism in primates. *Science*, **1984**, 225(4669), 1480-1482.
- [175] Magyar, K.; Szende, B. (-)-Deprenyl, a selective MAO-B inhibitor, with apoptotic and anti-apoptotic properties. *Neurotoxicology*, **2004**, 25(1-2), 233-242.
- [176] Tatton, W.; Chalmers-Redman, R.; Tatton, N. Neuroprotection by deprenyl and other propargylamines: glyceraldehyde-3-phosphate dehydrogenase rather than monoamine oxidase B. *J. Neural Transm.*, **2003**, 110(5), 509-515.
- [177] Baker, G. B.; Sowa, B.; Todd, K. G. Amine oxidases and their inhibitors: what can they tell us about neuroprotection and the development of drugs for neuropsychiatric disorders? *J. Psychiatry Neurosci.*, **2007**, 32(5), 313-315.
- [178] Sowa, B. N.; Holt, A.; Todd, K. G.; Baker, G. B. Monoamine oxidase inhibitors, their structural analogues, and neuroprotection. *Indian J. Exp. Biol.*, **2004**, 42(9), 851-857.
- [179] Song, M. S.; Baker, G. B.; Dursun, S. M.; Todd, K. G. The antidepressant phenelzine protects neurons and astrocytes against formaldehyde-induced toxicity. *J. Neurochem.*, **2010**, 114(5), 1405-1413.
- [180] MacKenzie, E. M.; Song, M.-S.; Dursun, S. M.; Tomlinson, S.; Todd, K. G.; Baker, G. B. Phenelzine: An old drug that may hold clues to the development of new neuroprotective agents. *Bull. Clin. Psychopharmacol.*, **2011**, 20, 179-186.
- [181] Wood, P. L.; Khan, M. A.; Moskal, J. R.; Todd, K. G.; Tanay, V. A.; Baker, G. Aldehyde load in ischemia-reperfusion brain injury: neuroprotection by neutralization of reactive aldehydes with phenelzine. *Brain Res.*, **2006**, 1122(1), 184-190.
- [182] Altar, C. A.; Whitehead, R. E.; Chen, R.; Wortwein, G.; Madsen, T. M. Effects of electroconvulsive seizures and antidepressant drugs on brain-derived neurotrophic factor protein in rat brain. *Biol. Psychiatry*, **2003**, 54(7), 703-709.
- [183] Balu, D.T.; Hoshaw, B.A.; Malberg, J.E.; Rosenzweig-Lipson, S.; Schechter, L.E.; Lucki, I. Differential regulation of central BDNF protein levels by antidepressant and non-antidepressant drug treatments. *Brain Res.*, **2008**, 1211, 37-43.
- [184] Yogeve-Falach, M.; Amit, T.; Bar-Am, O.; Youdim, M. B. The importance of propargylamine moiety in the anti-Parkinson drug rasagiline and its derivatives in MAPK-dependent amyloid precursor protein processing. *FASEB J.*, **2003**, 17(15), 2325-2327.
- [185] Maruyama, W.; Nitta, A.; Shamoto-Nagai, M.; Hirata, Y.; Akao, Y.; Youdim, M.; Furukawa, S.; Nabeshima, T.; Naoi, M. N-Propargyl-1 (R)-aminoindan, rasagiline, increases glial cell line-derived neurotrophic factor (GDNF) in neuroblastoma SH-SY5Y cells through activation of NF-kappaB transcription factor. *Neurochem. Int.*, **2004**, 44(6), 393-400.
- [186] Jenner, P. Preclinical evidence for neuroprotection with monoamine oxidase-B inhibitors in Parkinson's disease. *Neurology*, **2004**, 63(7 Suppl 2), S13-22.
- [187] Youdim, M. B.; Riederer, P. F. A review of the mechanisms and role of monoamine oxidase inhibitors in Parkinson's disease. *Neurology*, **2004**, 63(7 Suppl 2), S32-35.
- [188] Youdim, M. B.; Amit, T.; Falach-Yogeve, M.; Am, O. B.; Maruyama, W.; Naoi, M. The essentiality of Bcl-2, PKC and proteasome-ubiquitin complex activations in the neuroprotective-antiapoptotic action of the anti-Parkinson drug, rasagiline. *Biochem. Pharmacol.*, **2003**, 66(8), 1635-1641.
- [189] Magyar, K.; Haberle, D. Neuroprotective and neuronal rescue effects of selegiline: review. *Neurobiology (Bp)*, **1999**, 7(2), 175-190.
- [190] Berezovska, O.; Lleo, A.; Herl, L. D.; Frosch, M. P.; Stern, E. A.; Bacskai, B. J.; Hyman, B. T. Familial Alzheimer's disease presenilin 1 mutations cause alterations in the conformation of presenilin and interactions with amyloid precursor protein. *J. Neurosci.*, **2005**, 25(11), 3009-3017.
- [191] Ma, J.; Ito, A. Tyrosine residues near the FAD binding site are critical for FAD binding and for the maintenance of the stable and active conformation of rat monoamine oxidase A. *J. Biochem.*, **2002**, 131(1), 107-111.
- [192] Binda, C.; Hubalek, F.; Li, M.; Castagnoli, N.; Edmondson, D. E.; Mattevi, A. Structure of the human mitochondrial monoamine oxidase B: new chemical implications for neuroprotectant drug design. *Neurology*, **2006**, 67(7 Suppl 2), S5-7.
- [193] Edmondson, D. E.; Decolibus, L.; Binda, C.; Li, M.; Mattevi, A. New insights into the structures and functions of human monoamine oxidases A and B. *J. Neural Transm.*, **2007**, 114(6), 7032-705.
- [194] Grimsby, J.; Zentner, M.; Shih, J. C. Identification of a region important for human monoamine oxidase B substrate and inhibitor selectivity. *Life Sci.*, **1996**, 58(9), 777-787.
- [195] Tsugen, Y.; Ito, A. A key amino acid responsible for substrate selectivity of monoamine oxidase A and B. *J. Biol. Chem.*, **1997**, 272(22), 14033-14036.
- [196] Geha, R. M.; Chen, K.; Shih, J. C. Phe(208) and Ile(199) in human monoamine oxidase A and B do not determine substrate and inhibitor specificities as in rat. *J. Neurochem.*, **2000**, 75(3), 1304-1309.
- [197] Cao, X.; Rui, L.; Pennington, P. R.; Chlan-Fourney, J.; Jiang, Z.; Wei, Z.; Li, X. M.; Edmondson, D. E.; Mousseau, D. D. Serine 209 resides within a putative p38(MAPK) consensus motif and regulates monoamine oxidase-A activity. *J. Neurochem.*, **2009**, 111(1), 101-110.
- [198] Geha, R. M.; Chen, K.; Wouters, J.; Ooms, F.; Shih, J. C. Analysis of conserved active site residues in monoamine oxidase A and B and their three-dimensional molecular modeling. *J. Biol. Chem.*, **2002**, 277(19), 17209-17216.
- [199] Wu, H. F.; Chen, K.; Shih, J. C. Site-directed mutagenesis of monoamine oxidase A and B: role of cysteines. *Mol. Pharmacol.*, **1993**, 43(6), 888-893.
- [200] Vintem, A. P.; Price, N. T.; Silverman, R. B.; Ramsay, R. R. Mutation of surface cysteine 374 to alanine in monoamine oxidase A alters substrate turnover and inactivation by cyclopropylamines. *Bioorg. Med. Chem.*, **2005**, 13(10), 3487-3495.
- [201] Mitoma, J.; Ito, A. Mitochondrial targeting signal of rat liver monoamine oxidase B is located at its carboxy terminus. *J. Biochem. (Tokyo)*, **1992**, 111(1), 20-24.
- [202] Edmondson, D. E.; Mattevi, A.; Binda, C.; Li, M.; Hubalek, F. Structure and mechanism of monoamine oxidase. *Curr. Med. Chem.*, **2004**, 11(15), 1983-1993.
- [203] Zhuang, Z. P.; Marks, B.; McCauley, R. B. The insertion of monoamine oxidase A into the outer membrane of rat liver mitochondria. *J. Biol. Chem.*, **1992**, 267(1), 591-596.
- [204] Zhuang, Z.; Hogan, M.; McCauley, R. The *in vitro* insertion of monoamine oxidase B into mitochondrial outer membranes. *FEBS Lett.*, **1988**, 238(1), 185-190.
- [205] Baudhuin, P.; Beaufay, H.; Rahman-Li, Y.; Sellinger, O. Z.; Wattiaux, R.; Jacques, P.; De Duve, C. Tissue fractionation studies. 17. Intracellular distribution of monoamine oxidase, aspartate aminotransferase, alanine aminotransferase, D-amino acid oxidase and catalase in rat-liver tissue. *Biochem. J.*, **1964**, 92(1), 179-184.
- [206] Kroon, M. C.; Veldstra, H. Multiple forms of rat brain mitochondrial monoamine oxidase. Subcellular localization. *FEBS Lett.*, **1972**, 24(2), 173-176.
- [207] de Champlain, J.; Mueller, R. A.; Axelrod, J. Subcellular localization of monoamine oxidase in rat tissues. *J. Pharmacol. Exp. Ther.*, **1969**, 166(2), 339-345.
- [208] Erwin, V. G.; Deitrich, R. A. The labeling *in vivo* of monoamine oxidase by 14 C-pargyline: a tool for studying the synthesis of the enzyme. *Mol. Pharmacol.*, **1971**, 7(2), 219-228.

- [209] Muller, J.; Da Lage, C. Ultracytochemical demonstration of monoamine oxidase activity in nervous and non-nervous tissue of the rat. *J. Histochem. Cytochem.*, **1977**, *25*(5), 337-348.
- [210] Khuzhambardiev, M.; Kuz'mina, S. N.; Gorkin, V. Z.; Zbarskii, I. B. [Monoamine oxidase activity in cell nuclei and nuclear fractions of mouse liver under normal conditions and during development of ascitic sarcoma 37]. *Biull. Eksp. Biol. Med.*, **1972**, *73*(4), 37-39.
- [211] Gujrati, V. R.; Shanker, K.; Vrat, S.; Chandravati; Parmar, S. S. Novel appearance of placental nuclear monoamine oxidase: biochemical and histochemical evidence for hyperserotonergic state in preeclampsia-eclampsia. *Am. J. Obstet. Gynecol.*, **1996**, *175*(6), 1543-1550.
- [212] Ugun-Klusek, A.; Tamang, A.; Loughna, P.; Billett, E.; Buckley, G.; Sivasubramanian, S. Reduced placental vascular reactivity to 5-hydroxytryptamine in pre-eclampsia and the status of 5HT(2A) receptors. *Vascul. Pharmacol.*, **2011**, *55*(5-6), 157-162.
- [213] Student, A.K.; Edwards, D. J. Subcellular localization of types A and B monoamine oxidase in rat brain. *Biochem. Pharmacol.*, **1977**, *26* (24), 2337-2342.
- [214] Mousseau, D. D.; Baker, G. B.; Butterworth, R. F. Increased density of catalytic sites and expression of brain monoamine oxidase A in humans with hepatic encephalopathy. *J. Neurochem.*, **1997**, *68* (3), 1200-1208.
- [215] Fowler, J. S.; Alia-Klein, N.; Kriplani, A.; Logan, J.; Williams, B.; Zhu, W.; Craig, I. W.; Telang, F.; Goldstein, R.; Volkow, N. D.; Vaska, P.; Wang, G. J. Evidence that brain MAO A activity does not correspond to MAO A genotype in healthy male subjects. *Biol. Psychiatry*, **2007**, *62* (4), 355-358.
- [216] Cao, X.; Li, X. M.; Mousseau, D. D. Calcium alters monoamine oxidase-A parameters in human cerebellar and rat glial C6 cell extracts: possible influence by distinct signalling pathways. *Life Sci.*, **2009**, *85*(5-6), 262-268.
- [217] Kosenko, E. A.; Venediktova, N. I.; Kaminskii Iu, G. [Calcium and ammonia stimulate monoamine oxidase A activity in brain mitochondria]. *Izv. Akad. Nauk. Ser. Biol.*, **2003**, (5), 542-546.
- [218] Kabuto, H.; Yokoi, I.; Mori, A.; Murakami, M.; Sawada, S. Neurochemical changes related to ageing in the senescence-accelerated mouse brain and the effect of chronic administration of nimodipine. *Mech. Ageing Dev.*, **1995**, *80*(1), 1-9.
- [219] Fitzgerald, J. C.; Ufer, C.; De Girolamo, L. A.; Kuhn, H.; Billett, E. E. Monoamine oxidase-A modulates apoptotic cell death induced by staurosporine in human neuroblastoma cells. *J. Neurochem.*, **2007**, *103*(6), 2189-2199.
- [220] De Zutter, G. S.; Davis, R. J. Pro-apoptotic gene expression mediated by the p38 mitogen-activated protein kinase signal transduction pathway. *Proc. Natl. Acad. Sci. U S A*, **2001**, *98*(11), 6168-6173.
- [221] Wang, J.; Harris, J.; Mousseau, D. D.; Edmondson, D. E. Mutagenic probes of the role of Ser209 on the cavity shaping loop of human monoamine oxidase A. *Febs J.*, **2009**, *276*(16), 4569-4581.
- [222] Jiang, Y.; Chen, C.; Li, Z.; Guo, W.; Gegner, J. A.; Lin, S.; Han, J. Characterization of the structure and function of a new mitogen-activated protein kinase (p38 β). *J. Biol. Chem.*, **1996**, *271*(30), 17920-17926.
- [223] Lee, S. H.; Park, J.; Che, Y.; Han, P. L.; Lee, J. K. Constitutive activity and differential localization of p38 α and p38 β MAPKs in adult mouse brain. *J. Neurosci. Res.*, **2000**, *60*(5), 623-631.
- [224] Blanc, A.; Pandey, N. R.; Srivastava, A. K. Synchronous activation of ERK 1/2, p38mapk and PKB/Akt signaling by H2O2 in vascular smooth muscle cells: potential involvement in vascular disease (review). *Int. J. Mol. Med.*, **2003**, *11*(2), 229-234.
- [225] Konradi, C.; Riederer, P.; Youdim, M. B. Hydrogen peroxide enhances the activity of monoamine oxidase type-B but not of type-A: a pilot study. *J. Neural Transm. Suppl.*, **1986**, *22*, 61-73.
- [226] Blanc, A.; Pandey, N. R.; Srivastava, A. K. Distinct roles of Ca2+, calmodulin, and protein kinase C in H2O2-induced activation of ERK1/2, p38 MAPK, and protein kinase B signaling in vascular smooth muscle cells. *Antioxid. Redox. Signal.*, **2004**, *6* (2), 353-366.
- [227] Wu, J. B.; Shih, J. C. Valproic acid induces monoamine oxidase A via Akt/forkhead box O1 activation. *Mol. Pharmacol.*, **2011**, *80*(4), 714-723.
- [228] Kunduzova, O. R.; Bianchi, P.; Pizzinat, N.; Escourrou, G.; Seguelas, M. H.; Parini, A.; Cambon, C. Regulation of JNK/ERK activation, cell apoptosis, and tissue regeneration by monoamine oxidases after renal ischemia-reperfusion. *FASEB J.*, **2002**, *16*(9), 1129-1131.
- [229] Wong, W. K.; Ou, X. M.; Chen, K.; Shih, J. C. Activation of human monoamine oxidase B gene expression by a protein kinase C MAPK signal transduction pathway involves c-Jun and Egr-1. *J. Biol. Chem.*, **2002**, *277*(25), 22222-22230.
- [230] Bar-Am, O.; Amit, T.; Weinreb, O.; Youdim, M. B.; Mandel, S. Propargylamine containing compounds as modulators of proteolytic cleavage of amyloid- β protein precursor: involvement of MAPK and PKC activation. *J. Alzheimers Dis.*, **2010**, *21*(2), 361-371.
- [231] Tapia-Gonzalez, S.; Giraldez-Perez, R. M.; Cuartero, M. I.; Casarejos, M. J.; Mena, M. A.; Wang, X. F.; Sanchez-Capelo, A. Dopamine and α -synuclein dysfunction in Smad3 null mice. *Mol. Neurodegener.*, **2011**, *6*, 72.
- [232] Booysens, H. P.; Moraal, C.; Terre-Blanche, G.; Petzer, A.; Bergh, J. J.; Petzer, J. P. Thio- and aminocaffeine analogues as inhibitors of human monoamine oxidase. *Bioorg. Med. Chem.*, **2011**, *19*(24), 7507-7518.
- [233] Sloley, B. D.; Urichuk, L. J.; Morley, P.; Durkin, J.; Shan, J. J.; Pang, P. K.; Coutts, R. T. Identification of kaempferol as a monoamine oxidase inhibitor and potential neuroprotectant in extracts of Ginkgo biloba leaves. *J. Pharm. Pharmacol.*, **2000**, *52*(4), 451-459.
- [234] Dixon Clarke, S. E.; Ramsay, R. R. Dietary inhibitors of monoamine oxidase A. *J. Neural Transm.*, **2011**, *118*(7), 1031-1041.
- [235] Yoshino, S.; Hara, A.; Sakakibara, H.; Kawabata, K.; Tokumura, A.; Ishisaka, A.; Kawai, Y.; Terao, J. Effect of quercetin and glucuronide metabolites on the monoamine oxidase-A reaction in mouse brain mitochondria. *Nutrition*, **2011**, *27*(7-8), 847-852.